# SANT GADGE BABA AMRAVATI UNIVERSITY, AMRAVATI



# TWO YEAR POSTGRADUATE PROGRAMME

# IN ZOOLOGY

**FACULTY: SCIENCE AND TECHNOLOGY** 

M.Sc. II SEM III & IV (NEP)

(Courses effective from Academic Year 2024-25)

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# TWO YEAR POSTGRADUATE PROGRAMME M.Sc. ZOOLOGY under FACULTY: SCIENCE AND TECHNOLOGY Board of Studies in Zoology (Including Apiculture)

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# Sant Gadge Baba Amravati University Amravati FACULTY: SCIENCE AND TECHNOLOGY

#### Scheme of Teaching, Learning, Examination & Evaluation leading to Two Years PG Degree Master of Science (Zoology) following Three Years UG Programme w.e.f. 2023-24

(Two Years-Four Semesters Master's Degree Programme-NEP v23 with Exit and Entry Option

M.Sc. (Zoology) Second Year Semester-III [Level 6.5]

					Teaching & Learning Schen				Teaching & Learning Scheme				Duration		Examination & Evaluation Scheme						
G		T					of Maxin		num Mark	KS	1										
S. N.	Subject	Type of Course	Subject Code	T		ing P · We	Period ek	Credits			Hours	The	ory	P	ractical	Total	Mini	Minimum Passing			
				L	Т	P	Total	L/T	Practical	Total		Theory +MCQ Internal	Theory External	Internal	External	Marks	Marks Internal	Marks External	Grade		
1	DSC-I.3 Molecular Cytogenetics	Th-Major	3 ZOO 1	4	-	-	4	4		4	3	40	60			100	16	24	P		
2	DSC-II.3 Population, Quantitative and Evolutionary Genetics	Th-Major	3 ZOO 2	4	-	-	4	4		4	3	40	60			100	16	24	P		
3	DSC-III.3 Endocrinology	Th-Major	3 ZOO 3	3	-	-	3	3		3	3	40	60			100	16	24	P		
4	DSE-III: Entomology-I/ Fishery -I/ Animal physiology-I/ Molecular Biology-I	Th-Major Elective	3 ZOO 4	3	-	-	3	3		3	3	40	60			100	16	24	P		
																		n Passing arks			
5	DSC I.3 and DSC II.3.(Lab 5)	Pr-Major	3 ZOO 5	-	-	4	4		2	2	6			50	50	100		50	P		
6	DSC-III.3.and DSE III (Lab 6)	Pr-Major	3 ZOO 6	•	-	4	4		2	2	6			50	50	100		50	P		
7	Research Project Phase-I	Major		•	2	4	6	2	2	4				50		50	2	25	P		
8	Co-curricular Courses: Health and wellness, Yoga Education, Sports and fitness, Cultural Activities, NSS/ NCC, Fine / Applied/ Visual/ Performing Arts During Semester I, II, III and IV	Generic <b>Optional</b>		Cun Fro	) Hou nulati om SE SEM	ively M I															
	TOTAL				ı	1				22						(50					
	TOTAL 22			1				650													

#### L: Lecture, T: Tutorial, P: Practical / Practicum

Pre-requisite Course mandatory if applicable: **Prq**, Theory: **Th**, Practical/Practicum: **Pr**, Faculty Specific Core: **FSC**, Discipline Specific Elective: **DSE**, Laboratory: **Lab**, **OJT**: On Job Training: Internship/ Apprenticeship; Field projects: **FP**; **RM**: Research Methodology, Research Project: RP, **Co-curricular Courses: CC**.

Note: Co-curricular Courses: In addition to the above, CC also include but not limited to Academic activities like paper presentations in conferences, Aavishkar, Startups, Hackathon, Quiz competitions, Article published, Participation in Summer school /Short term course, Scientific Surveys, Societal Surveys, Field Visits, Study tours, Industrial Visits, online /offline Courses on Yoga (Yoga for Ego development, Yoga for Ego develo

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Scheme of Teaching, Learning, Examination & Evaluation leading to Two Years PG Degree Master of Science (Zoology) following Three Years UG Programme w.e.f. 2023-24

(Two Years-Four Semesters Master's Degree Programme-NEP v23 with Exit and Entry Option

M.Sc. (Zoology) Second Year Semester-IV [Level 6.5]

			Teaching & Learning Scheme Duration			Teaching & Learning Scheme							Examination & Evaluation				Scheme			
								of		Maxi	imum Marks									
S.	N. Subject	Type of Course	Subject Code	Т		ing P r We	Period ek	(	Credits		Exam Hours	Theo	ory	P	ractical	Total	Minimum Pa		ssing	
				L	Т	P	Total	L/T	Practical	Total		Theory +MCQ Internal	Theory External	Internal	External	Marks	Marks Internal	Marks External	Grade	
	DSC-I.4 Biochemistry	Th-Major	4 ZOO 1	4	-	-	4	4		4	3	40	60			100	16	24	P	
	DSC-II.4 Enzymology and Biostatistics	Th-Major	4 ZOO 2	4	-	-	4	4		4	3	40	60			100	16	24	P	
	DSC-III.4 General Parasitology	Th-Major	4 ZOO 3	3	-	-	3	3		3	3	40	60			100	16	24	P	
,	DSE-IV: Entomology-II/ Fishery -II/ Animal physiology-II/ Molecular Biology-II/	Th-Major Elective	4 ZOO 4	3	-	-	3	3		3	3	40	60			100	16	24	P	
																		n Passing arks		
	DSC I.4 and DSC II.4.(Lab 7)	Pr-Major	4 ZOO 5	-	-	4	4		2	2	6			50	50	100	5	50	P	
	DSC-III.4.and DSE IV (Lab 8)	Pr-Major	4 ZOO 6		-	4	4		2	2	6			50	50	100	5	50	P	
,	Research Project Phase-II	Major		-	2	8	10	2	4	6	3			75	75	150	7	<b>'</b> 5	P	
	Co-curricular Courses: Health and wellness, Yoga Education, Sports and fitness, Cultural Activities, NSS/ NCC, Fine / Applied/ Visual/ Performing Arts During Semester I, II, III and IV	Generic <b>Optional</b>		Cun Fro	) Hou nulat m SE SEM	ively EM I														
	TOTAL									24						750				

#### L: Lecture, T: Tutorial, P: Practical / Practicum

Pre-requisite Course mandatory if applicable: **Prq**, Theory: **Th**, Practical/Practicum: **Pr**, Faculty Specific Core: **FSC**, Discipline Specific Elective: **DSE**, Laboratory: **Lab**, **OJT**: On Job Training: Internship/ Apprenticeship; Field projects: **FP**; **RM**: Research Methodology, Research Project: RP, **Co-curricular Courses: CC**.

Note: Co-curricular Courses: In addition to the above, CC also include but not limited to Academic activities like paper presentations in conferences, Aavishkar, Startups, Hackathon, Quiz competitions, Article published, Participation in Summer school /Short term course, Scientific Surveys, Societal Surveys, Field Visits, Study tours, Industrial Visits, online /offline Courses on Yoga (Yoga for Ego development, Yoga for Ego develo

### Sant Gadge Baba Amravati University, Amravati Name of the Programme: M.Sc. (SEM-III) Subject: Zoology (NEPv23)

#### POs:

The post graduate course of Zoology will provide theoretical as well as experimental knowledge as per the courses included under the syllabi by which build up creativity in students will lead towards thorough learning and development of ideas of research work and will become ready to face recent challenges. Students can attain the employability skills through the experiences based on their practical knowledge.

#### After completion of MSc in Zoology successfully, the students would be able to.....

- 1. Demonstrate the significance of the topics of syllabi and evaluate its relevance. Think creatively for its gravity and develop ideas.
- 2. Interpret scientific ideas and can do its analysis. Create experiments independently and draw inferences by sharing it with others.
- 3. Derive information from various digital sources. Develop skills for scientific writing and present the data and analyse it scientifically.
- 4. Articulate scientific ideas lay down a hypothesis; design the pathway to develop research ideas.
- 5. Acquaint skills in handling the instruments and different techniques through practicals and developing the scientific temperaments for research.
- 6. Develop competence through healthy atmosphere and quality intercommunication with different groups.
- 7. Understand environmental and sustainability issues and its sensitivity in relation to regional relevance.
- 8. Get the facility of different training and internship programs through job-oriented curriculum
- 9. Utilize the sources confidently and independently and develop self-sustenance.

#### **PSOs:**

### Upon completion of the Programme successfully, students would be able to .......

- 1. Learn to prepare the checklist and inventories through the identification of the fauna in local areas being Melghat Tiger Reserve and Pohra Forest are very nearer to survey.
- 2. Gain comprehensive knowledge about different animals and develop confidence to handle them during research work
- 3. Interpret metabolic pathways, their correlation in concern with prokaryote and eukaryotes.
- 4. Compare genetic aspects, genetic traits, diseases and their specific causes.
- 5. Survey and analyse data of the various kinds of diseases in the locality.
- 6. Understand various strategies and phenomena related to animal reproduction and their development.
- 7. Get acquainted with conservation strategies and environmental threats to reduce and save energy through Wildlife Week Celebration.
- 8. Compare the different developmental events during embryogenesis of different animals.

# EMPLOYABILITY POTENTIAL

- After Completion of this course Field Biologist or Ecologist in different govt. establishments like NTCA, Division Office, Wildlife Wings Wildlife Consultant EIA Report Designing Firm,
- Wildlife Conservation Educator in NGOs like WWF, WCT, WTI, Conservation Geneticist in research organisations,
- GIS Specialist in Govt Establishments, Environmental Impact Assessment (EIA) Specialist: Assessing the potential impact of development Projects on wildlife and proposing mitigation measures.

- Wildlife Forensics Specialist in research labs
- Research Associate or Assistant: Assisting senior researchers in wildlife conservation projects and Data analysis.
- Wildlife Photographer or Filmmaker: Using visual media to raise awareness about wildlife Conservation and natural habitats.
- Wildlife Tour Guide or Naturalist: Conducting guided tours in wildlife sanctuaries, national parks, or Eco-tourism sites.
- Wildlife Project Manager: Overseeing and coordinating conservation projects aimed at protecting Specific wildlife species or habitats.
- Environmental Educator: Developing and delivering educational programs related to wildlife Conservation and environmental protection.
- Environmental Scientists, Ecologist, Environmental manager, scientific technical writer, zoo manager, Lab technician

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### **Examination, Evaluation and Assessment Scheme**

#### 1. External Theory Examination - 60 Marks (Minimum Passing Marks = 24)

Question Paper Pattern (for example)							
Unit No.	Question No.	Question Type for Unit	Marks				
with either OR, from	estions of 10 mark each (any Three units out of le shown in Question	<ul><li>a) Long answer question.</li><li>OR</li><li>b) Long answer question.</li></ul>	10 10				
mark each with eithe	answer questions of 05 or OR, from (any three ) as example shown in	<ul> <li>c) Short answer question.</li> <li>d) Short answer question.</li> <li>OR</li> <li>e) Short answer question.</li> <li>f) Short answer question.</li> </ul>	05 05 05 05				

### 2. Internal Assessment Marks (Theory) - 40 Marks

- Internal assessment marks (Theory) shall be based on two class tests (20 marks each), student will have to score at least 40 % Marks, that is, 16 Marks for passing the theory internal course.
- In case, even after completing the requisite term-work, the student is unable to score minimum prescribed marks in Internal Theory Examination, that is, 16, he/she will be declared as Fail.
- Now, MCQ, if any, will be part of the Internal Assessment only.
- However, the concerned college/institute/university department shall give one more opportunity to such failure students.
- Thus, failure students will get another chance to clear their theory courses/subjects.
- The remedial re-examination of such failure students shall be conducted before the commencement of end-semester university examinations, so that the concerned college/institute/university department can submit the revised internal marks of such failure students to the University in due course of Time as instructed by the university.

# M. Sc. Zoology (NEPv23) Semester-III

Course: DSC I.3 Molecular Cytogenetics Th- Major						
Subject Code: 3 ZOO 1	No. of Credits: 4	No. of hours per week: 4				
Exam duration: 3 Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 60				

# COs: 3 ZOO 1

- 1. Students acquire knowledge about the mutation and different causes of mutation.
- 2. The student gain in-depth knowledge regarding transposition mechanisms in prokaryotes and eukaryotes, and their significance.
- 3. Special features of microbial genetics, and organelle genome, their replication and mapping.
- 4. The student gain an in depth knowledge regarding the events and regulation of cell cycle, its alteration and causes of cancer. Genes involved in the regulation of cancer.

Unit	Contents	Hours
	Mutation:	
	1.1 Mutation induced by chemicals and radiation.	
	1.2 Mutation caused by DNA replication machinery.	10
Unit-I	1.3 Hot spots of mutation.	10
	1.4 Detection of mutagens-The Ames Test.	
	1.5 DNA repair mechanisms.	
	Somatic Cell Hybridization:	
	2.1 Agents of cell fusion, Mechanism of cell fusion, Selection of hybrids.	
	2.2 Radiation hybrid panels and gene mapping.	
Unit-II	Epigenetics:	10
	2.3 Mechanisms of histone modification.	
	2.4 Prions and epigenetic inheritance.	
	2.5 Polycomb mechanisms and epigenetic control of gene activity.	
	Transposable Genetic Elements:	
	3.1Transposable elements in bacteria, <i>Drosophila</i> and humans.	
	Genetics of Cancer:	
Unit-III	3.2 Properties of cancer cells.	10
	3.3 Metastasis.	
	3.4 Oncogenes.	
	3.5 Tumor suppressor genes.	
	Human Genetic Diseases and Disorders:	
	4.1 Diseases Caused Due to Numerical Abnormalities of Chromosomes:	
	Edwards syndrome, Down syndrome, Turner syndrome, Klinefelter syndrome.	
	4.2 Diseases Caused Due to Structural Abnormalities of Chromosomes:	
	Cri-du-chat syndrome, Prader-Willi syndrome.	
Unit-IV	4.3 Human Metabolic Disorders:	10
	Phenylketonuria, Lesch-Nyhan syndrome, Tay-Sachs disease,	
	4.4 Other Genetic Diseases: Duchenne muscular dystrophy, Sickle cell anemia,	
	Thalassemia, Alzheimer's disease, Parkinson's disease.	
	4.5 Diseases Caused Due to Defects in Mitochondrial DNA.	
	Bacterial Genetics:	
Unit-V	5.1 Bacterial transformation, conjugation and transduction.	10

	Drosophila Genetics:					
	5.2 <b>Polytene Chromosomes:</b> Polytenization process and its significance, Regulation of					
	puffing activity.					
	5.3 <b>Behavioral Genetics:</b> Mutants, Behavioural traits, Genetic and molecular basis of					
	biological rhythm.					
	Extra-chromosomal Inheritance:					
	5.4 Maternal inheritance of <i>kappa</i> particles in <i>Paramecium</i> , Shell coiling in <i>Lymnaea</i> .					
	Molecular Cytogenetic Techniques:					
	6.1 Human karyotyping.					
** ** ***	6.2 Flow cytometry (for chromosome separation).					
Unit-VI	6.3 Chromosome painting.	10				
	6.4 Fluorescence in situ hybridization (FISH).					
	6.5 <b>DNA Fingerprinting:</b> Principle, procedure and applications.					
l		1				

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- 5. Brown, T. A. Genetics-A Molecular Approach, 3/Ed. Garland Science.
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- 23. Snustad, D. Peter and M.J. Simmons. Principles of Genetics, 7/Ed. Wiley-Blackwell.
- 24. Strachan Tom and Andrew Read. Human Molecular Genetics, 4/Ed. Garland Science.
- 25. Strickberger, M. W. Genetics. Pearson Education, Inc.
- 26. Tollefsbol, T. Handbook of Epigenetics: The New Molecular and Medical Genetics.
- 27. Turnpenny Peter D. et al. Emery's Elements of Medical Genetics, 15/Ed.
- 28. Vogel, F. and A.G. Motulsky. Human Genetics. Springer-Verlog, NY
- 29. Weaver, R.F. & P. W. Hedrick: Genetics 3/Ed. Wm. C. Brown Pub. London.
- 30. Wright Alan and Nicholas Hastie. Genes and Common Diseases. Cambridge University

Course: DSC II.3 Population, Quantitative and Evolutionary Genetics Th- Major						
Subject Code: 3 ZOO 2	No. of Credits: 4	No. of hours per week: 4				
Exam duration: 3 Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 60				

### COs: 3 ZOO 2

- 1. Student understand the concept of Genetic Structure of Populations.
- 2. Gaining knowledge of forces changing the gene frequency.
- 3. Learn methods of construction of phylogenetic tree and methods to retrieve the nucleotide sequences.
- 4. Understand the quantitative genetics and several factors influencing the quantitative traits.

Population Genetics: Genetic Structure 1.1 Genotype frequencies. 1.2 Allele frequencies. 1.3 Hardy—Weinberg principle. 1.4 Applications of Hardy-Weinberg principle. 1.5 Exceptions to Hardy-Weinberg principle. 1.6 Consanguineous mating and its consumption of the principle. 2.1 Nonrandom mating.	inciple. ciple. sequences.	10					
1.2 Allele frequencies.  1.3 Hardy—Weinberg principle.  1.4 Applications of Hardy-Weinberg principle.  1.5 Exceptions to Hardy-Weinberg principle.  1.6 Consanguineous mating and its considered principle.  Population Genetics: Forces Changing	ciple. sequences.	10					
Unit-I  1.3 Hardy—Weinberg principle.  1.4 Applications of Hardy-Weinberg principle.  1.5 Exceptions to Hardy-Weinberg principle.  1.6 Consanguineous mating and its consumption of the	ciple. sequences.	10					
1.4 Applications of Hardy-Weinberg pr 1.5 Exceptions to Hardy-Weinberg prin 1.6 Consanguineous mating and its cons  Population Genetics: Forces Changin	ciple. sequences.	10					
1.5 Exceptions to Hardy-Weinberg prin 1.6 Consanguineous mating and its cons  Population Genetics: Forces Changin	ciple. sequences.						
1.6 Consanguineous mating and its cons  Population Genetics: Forces Changin	sequences.						
Population Genetics: Forces Changin							
	g Gene Frequencies-						
2.1 Nonrandom mating.							
2.2 Mutation.							
Unit-II 2.2 Migration.		10					
2.3 Genetic drift.	2.3 Genetic drift.						
2.4 Natural selection.							
2.5 Balance between mutation and select	ction.						
<b>Quantitative Genetics:</b>							
3.1 Quantitative traits.							
	3.2 Factors influencing quantitative traits.						
Unit-III 3.3 Molecular analysis of quantitative to	3.3 Molecular analysis of quantitative trait loci.						
3.4 Genotype-environmental interaction	ns.						
3.5 Inbreeding depression and heterosis							
Evolution:							
4.1 Theories of Evolution: Lamarck's the	neory, Neo-Lamarckism, Darwin's theory,						
	lutation theory, Modern synthetic theory.	4.0					
Unit-IV 4.2 Origin of basic organic monomers a	and polymers, Miller-Urey experiment.	10					
4.3 Origin of organized structures-coace	ervates and microspheres.						
4.4 Geological time-scale.							
<b>Evolutionary Genetics:</b>							
5.1 Genetic variation in natural populat	ions.						
5.2 Nucleotide substitution in DNA seq	uences.						
Unit-V 5.3 Rates of nucleotide substitutions.		10					
5.4 Variation in evolutionary rates betw	veen genes.						
5.5 Rates of evolution in mitochondrial	DNA.						
5.6 Molecular clocks.							

	Molecular Phylogenetics:	
	6.1 Features of phylogenetic trees.	
	6.2 Methods of phylogenetic tree reconstruction.	
	6.3 Nucleic acid phylogeny based on DNA-DNA hybridization, Restriction	
Unit-VI	Enzyme sites, and Nucleotide sequence comparisons and homologies.	10
	6.4 Acquisition and Origin of New Genes: Multigene families, Gene duplication	
	and gene conversion.	
	6.5 Protein phylogeny.	

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- 3. Futuyma, Douglas J. and Mark Kirkpatrick. Evolution, 4/Ed. Sinauer Associates, Inc.
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- 7. Hartl, D. L. Essential Genetics and Genomics, 7/Ed. Jones & Bartlett Pub.
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- 12. Lewis Ricki. Human Genetics-Concepts and Applications, 12/Ed. McGraw-Hill Education.
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- 17. Russell, P. J. iGenetics, 3/Ed. Pearson Education, Inc.
- 18. Smith, John Maynard. Evolutionary Genetics, 2/Ed.Oxford University Press.
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- 20. Strachan Tom and Andrew Read. Human Molecular Genetics, 4/Ed. Garland Science.
- Verma, P. S. and V. K. Agarwal. Cell Biology, Genetics, Molecular Biology, Evolution and Ecology. S. Chand and Company Ltd.
- 22. Vogel, F. and A.G. Motulsky. Human Genetics. Springer-Verlog, NY.

Course: Pr- Major Laboratory 5								
(DSC I.3- Molecular Cytogenetics + DSC II.3 - Population, Quantitative and Evolutionary Genetics)								
Subject Code: 3 ZOO 5	No. of Credits: 2	No. of hours per week: 4						
Exam duration: 6 Hrs	Maximum Marks: 100 (External:50 Internal:50)	Total No. of contact hours: 60						

#### A) Practicals for Molecular Cytogenetics

- 1. Demonstration of Barr bodies in leucocytes of human female.
- 2. Demonstration of salivary gland chromosomes from Chironomous/Drosophila larvae.
- 3. Study of mitosis in cleaving eggs of Frog/any invertebrate.
- 4. Histological demonstration of meiosis in mammalian testis.
- 5. Preparation of human karyotype using photograph/picture.
- 6. Culture of Drosophila and study of life cycle and sexual polymorphism.
- 7. Identification of wing and eye mutants in Drosophila.
- 8. Extraction of DNA.
- 9. Estimation of DNA (spectrophotometric/colorimetric).
- 10. Extraction of RNA.
- 11. Estimation of RNA (spectrophotometric/colorimetric).
- 12. Problems on Genetics based on dihybrid crosses, sex-linked inheritance and blood groups.
- 13. Study of human genetic traits.

# B) Practicals for Population, Quantitative and Evolutionary Genetics:

- 1. Problems based on calculation of gene/allele frequencies using Hardy-Weinberg Principle.
- 2. Retrieval of DNA/protein sequences from databases.
- 3. Biological sequence editing.
- 4. Sequence alignment: database searches (BLAST, FASTA).
- 5. Phylogenetic tree construction.
- 6. Analysis of phylogenetic tree.

### The examinee shall be required to produce at the practical examination the following:

Practical record book duly signed by teacher in-charge and certified by the Head of the Department as a bonafide work of the examinees.

### DISTRIBUTION OF PRACTICAL MARKS

#### **External Practical Marks**

#### **Experiments from Section A)** Molecular Cytogenetics

1. Estimation/Experiment	10 marks
2. Cytological Preparation	05 marks
3. Problem on Genetics	05 marks

# Experiments from Section B) Population, Quantitative and Evolutionary Genetics

4.	DNA/protein	sequence	alignment	PT	tree construction.	10 Marks
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5. Retrieval of DNA/protein sequences from database......05 Marks

TOTAL 50 Marks

#### **Internal Practical Marks**

 1. Certified Practical Record
 : 20 marks

 2. Submission of Stained Permanent sides
 : 10 marks

 3. Submission of phylogenetic tree
 : 10 marks

 4. Student Performance
 : 10 marks

TOTAL 50 Marks

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Course: DSC III.3 Endocrinology Th- Major			
Subject Code: 3 ZOO 3	No. of Credits: 4	No. of hours per week: 4	
Exam duration: 3 Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 60	

# COs: 3 ZOO 3

- **1.** Describe types of hormones and their action at cellular and genetic level
- **2.** Identify the type of hormone on the basis of its action.
- **3.** Identify various parts of important endocrine glands and describe the role of their hormones.
- **4.** Describe hormonal interplay during metamorphosis

Unit	Contents	Hours	
	Anatomy, Histology and Hormones of Vertebrate Endocrine Glands:		
<b>T</b> T *4 <b>T</b>	1.1 Pituitary, Thyroid, Parathyroid, Adrenal, Pineal gland and Islets of Langerhans.		
	1.2 Endocrine placenta, testis and ovary.	10	
Unit-I	1.3 Ultimobranchial glands.	10	
	1.4 Urophysis and Corpuscles of Stannius in fishes.		
	1.5 Hormones from hypothalamus.		
	Classification and Biosynthesis of Hormones:		
	2.1 Evolution of hormones.		
	2.2 Classification of Hormones (peptides, steroids and amino acid derived).		
Unit-II	2.3 Biosynthesis of steroid hormones.	10	
	2.4 Biosynthesis of T3, T4 and epinephrine.		
	2.5 Biosynthesis of peptide hormones.		
	2.6 Hormone secretion, transport, and clearance from the Blood.		
	Action and Functions of Hormones:		
	3.1 Hormone action at membrane level (Ex. Insulin).		
	3.2 Hormone action at genetic level (Ex. Testosterone).		
T TT	3.3 Hormones in digestion.	10	
Unit-III	3.4 Hormonal regulation of carbohydrate, lipid and protein metabolism.	10	
	3.5 Hormonal regulation of growth and reproduction.		
	3.6 Other hormones and their Functions: Renin, Angiotensin,		
	Atrial Natriuretic Factor (ANF), and Erythropoietin.		
	Endocrine Disorders:		
	4.1 Thyroid hormones and disorders.		
	4.2 Parathyroid hormones and disorders.		
TT *4 TT7	4.3 Pituitary hormones and disorders.	10	
Unit-IV	4.4 Adrenal hormones and disorders.	10	
	4.5 Endocrine changes in pregnancy.		
	4.6 <b>Diabetes:</b> Causes of Type I and Type II Diabetes, Effects of Diabetes		
	(Retinopathy, Neuropathy and Nephropathy), Gestational Diabetes.		
	Therapeutic and Behavioral Endocrinology:		
Unit-V	5.1 Hormone replacement therapy.		
	5.2 Risks and benefits of hormone replacement therapy.	10	
	5.3 Transfeminine and transmasculine hormone therapy	10	
	5.4 Hormones and social behavior.		
	5.5 Hormones and parental behaviour.		

	Other Aspects of Endocrinology:	
	6.1 Effect of athletic performance on hormonal systems.	
TI 14 T/T	6.2 Performance-enhancing (Ab) use of hormones.	10
Unit-VI	6.3 Adipose tissue as an endocrine organ.	10
	6.4 Hormonal control of Crustacean metamorphosis.	
	6.5 Hormonal control of Insect metamorphosis.	

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- 2. Dattani Mehul T. (Editor). Brook's Clinical Pediatric Endocrinology, 7/Ed. Wiley.
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- 4. Gardner David G and Dolores Shoback. Basic and Clinical Endocrinology, 9/Ed. McGraw Hill Medical.
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- 10. Ilie, Ioana R. Introduction to Endocrinology. Springer.
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- 19. Norman Lavin. Manual of Endocrinology and Metabolism, 5/Ed. Wolters Kluwer.
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- 26. Strauss, Jerome F. and Robert L. Barbieri. Reproductive Endocrinology. Elsevier.

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Course: DSE III Entomology-1 Th- Major Elective-I		
(Insect Classification, Morphology and Physiology)		
Subject Code: 3 ZOO 4	No. of Credits: 3	No. of hours per week: 3
Exam duration: 3 Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 45

# CO's: 3 ZOO 4

- 1: Students are trained in the basics of insect classifications.
- 2: Insect anatomy of various insects are studied in detail.
- 3: Insect physiology of various insects are studied in detail.
- 4: Acquire an understanding of insect metamorphosis process.

Units	Syllabus Content	Hours	
	Insect classification		
	1.1 Classification of class insecta (class, subclass, orders with examples).		
	1.2 Distinguishing Characters and general biology of Apterygota and Pterygota.		
Unit I	1.3 Distinguishing Characters and general biology of endopterygota and exopterygota.	7	
	1.4 Distinguishing Characters and general biology of coleoptera and diptera.		
	1.5 Distinguishing Characters and general biology of orthoptera and hemiptera.		
	Structure of Insects:		
	2.1 Structure of insect body wall.		
	2.2 Insect Head- structure and modification.		
Unit II	2.3 Types of mouthparts and antennae.	8	
	2.4 Thorax- Areas and sutures of tergum.		
	2.5 Wings: structure and venation.		
	2.6 Legs: structure and modifications		
	Segmentation and Receptors:		
	3.1 Abdomen- Segmentation and appendages.		
	3.2 Genitalia and their modifications.		
Unit III	3.3 Insect sense organs –mechanoreceptors.	7	
	3.4 Photoreceptors.		
	3.5 Chemoreceptors.		
	Digestive and Circulatory System of Insects:		
	4.1 Structure of Digestive system of insects.		
	4.2 Physiology of digestive system.		
Unit IV	4.3 Structure of Circulatory system of insects.	8	
	4.4 Physiology of Circulatory system		
	4.5 Modifications of Circulatory system.		
	Respiratory and Excretory System of Insects:		
	5.1 Structure of respiratory system of insects.		
	5.2 Physiology of respiratory system.		
Unit V	5.3 Structure of excretory system of insects.	8	
	5.4 Physiology of excretory system.		
	5.5 Modifications of excretory system.		

	Physiology of Metamorphosis:.	
	6.1 Physiology of insect growth.	
	6.2 Physiology of metamorphosis in insects.	_
Unit VI	6.3 Polyphenism and diapause.	7
	6.4 Biochemistry of insect cuticle	
	6.5 Physiology of moulting process.	

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- 2. David BV & Ananthkrishnan TN. 2004. General and Applied Entomology.
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Course: Pr- Major Laboratory 6		
(DSC III.3 Endocrinology + DSE III Entomology-1 Insect Classification, Morphology and Physiology)		
Subject Code: 3 ZOO 6	No. of Credits: 2	No. of hours per week: 4
Exam duration: 6 Hrs	Maximum Marks: 100	Total No. of contact hours: 60
	(External:50 Internal:50)	

# A) Practicals for Endocrinology

- 1. Histological study of various vertebrate endocrine glands using permanent slides / photographs (Mammals, Crab and Insects).
- 2. Histology of various vertebrate endocrine glands (microtomy).
- 3. Morphological and histological study of various insect neuroendocrine structures.
- 4. Effect of toxicants on histoarchitecture of various endocrine glands.
- 5. Study of various endocrine disorders using photographs.
- 6. Study of effect of exogenous growth hormone on the growth of fish.
- 7. Study of vaginal smear during oestrous cycle of Rat.

#### B) Practicals for Entomology-1 Insect Classification, Morphology and Physiology

- 1. Identification and classification of insects from different orders (2 example from each order)
- 2. Mounting of different mouth parts of Mosquito, House fly, Cockroach.
- 3. Mounting of different antennae from Mosquito, Cockroach and other pest insects.
- 4. Haemolymph collection, staining and identification of haemocytes.
- 5. Slide preparation for different types of insect wings.
- 6. Study of different insect wing venation pattern.
- 7. To study the insect legs and their modifications
- 8. To study the types of insect larvae and pupae.
- 9. Study of Digestive system of insects by using chart/ PPT/ E-content.
- 10. Study of Circulatory system of insects by using chart/ PPT/ E-content.
- 11. Study of Excretory System of Insects by using chart/ PPT/ E-content.
- 12. Study of Respiratory system of insects by using chart/ PPT/ E-content.
- 13. Study of life cycle of House fly, Cockroach, Mosquito.

# The examinee shall be required to produce at the practical examination the following:

Practical record book duly signed by teacher in-charge and certified by the Head of the Department as a bonafide work of the examinees.

# DISTRIBUTION OF PRACTICAL MARKS External Practical Marks

### **Experiments from Section A) Endocrinology**

1. Histological Preparation/Experiment	12 Marks
2. Identification, Labeling and Comment on	
Spots / Photographs of endocrine disorders	08 Marks

#### Experiments from Section B) Entomology-1 Insect Classification, Morphology and Physiology

3. Identification and classification of insects (06 spots)	.12 Marks
4. Temporary slide preparation	08 Marks
6. Viva Voce	10 Marks

TOTAL 50 Marks

#### **Internal Practical Marks**

1. Certified Practical Record	: 20 Marks
2. Submission of Stained Permanent sides	: 10 Marks
3. Submission of photographic collection of insect pest	: 10 Marks
4. Student Performance	: 10 Marks

TOTAL 50 Marks

Course: DSE III Fishery-I Th- Major Elective-I Fresh Water Fish Culture			
Subject Code: 3 ZOO 4 No. of Credits: 3 No. of hours per week: 3			
Exam duration: 3 Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 45	

# COs

- 1. Students will get acquaint with the various components of fish farm.
- 2. Students will get practical knowledge on feed formulation
- 3. Students will gain knowledge about different types of fish breeding techniques.
- 4. Students will gain knowledge about different types of fish pathogen and their treatments.

Unit	Course Content	Hours
	1.1 Riverine fisheries: Major river system in India.	
	1.2 Fishing methods, representative ichthyofauna, and recent catch statistics.	
Unit I	1.3 Problems encountered in fisheries development of major rivers.	07
	1.4 Cold water fisheries: Cold water fisheries resources of India.	
	1.5 Representative species of fishes of cold water bodies of India.	
	2.1 Reservoir and Lacustrine fisheries: Definition and ecological features of	
	reservoirs and lakes.	
	2.2 Major reservoirs and lakes in India with emphasis on capture fisheries.	
Unit II	2.3 Development of reservoir fisheries in India.	07
	2.4 Estuarine fisheries: Definition and classification of estuaries, capture fisheries	
	resident and migrant species, fishing methods.	
	2.5 Fisheries of brackish water lake and backwaters.	
	3.1 Scope and present status of aquaculture: Principles of site selection of various	
	kinds of fish farms- quality and productivity of water, soil characteristics and	
	other parameters.	
	3.2 Carp Culture: Pre -stocking, Stocking and Post stocking management of	
	Nursery, Rearing and Stocking pond.	0.0
Unit III	3.3 Criteria for selection of species for culture.	08
	3.4 Different systems of aquaculture: Monoculture, Polyculture, Integrated fish	
	farming, cage culture, pen culture.	
	3.5 Aquaculture diversification- Aquaponics system, Biofloc culture, IMTA and	
	periphyton culture	
	4.1 Reproductive Biology of Fishes: Morphology and Histology of Pituitary, Testis	
	and Ovary.	
	4.2 Hormonal regulation of spermatogenesis and oogenesis.	
	4.3 Role of GnRH and GTH on gonadal functions.	0.0
Unit IV	4.4 Natural breeding of fishes. Induced Breeding of Indian major carps and exotic	08
	carps by Hypophysation.	
	4.5 Hatchery Technology for Indian Major Carps. Glass jar hatchery, Chinese	
	hatchery and other hatchery systems.	
	5.1 Food and feeding habits of freshwater fishes, prawn, mussel and oysters.	
	5.2 Fundamental of Fish nutrition: Principles of fish nutrition and terminologies,	
Unit V	5.3 Role of nutrients and their requirement: amino acids, fatty acids, proteins,	08
	lipids, carbohydrates, vitamins and minerals for various growth stages of	
	freshwater carp, prawn and mussel.	
	1	

	5.4 Feed formulation: Conventional and non-conventional feed stuffs, feed	
	formulation technology, growth promoting agents in aqua feed, single cell	
	protein (SCP).	
	5.5 Presence of anti-nutritional factors and their removal procedures.	
	Supplementary feed: Kind, Composition and nutrient source.	
	6.1 Fish pathology and Management, Pathological processes: Cellular response to	
	injury, inflammatory response to diseases,	
	6.2 Role of stress in fish disease, Parasitic and non-parasitic diseases: Protozoan	
	diseases, fungal diseases, bacterial diseases, viral diseases.	
Unit VI	6.3 Disease surveillance and reporting, Disease control through environmental	07
	management, Sanitary and phytosanitary agreement,	
	6.4 Vaccines used in aquaculture, Immuno-stimulants used in aquaculture,	
	Bioremediation.	

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- 27. Ujwala Jadhav (2010): Aquaculture Technology and Environment. Publ. PHI Publication

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Course: Pr- Major Laboratory 6			
(DSC III.3 Endocrinology + DSE III Fishery-I Fresh Water Fish Culture)			
Subject Code: 3 ZOO 6 No. of Credits: 2 No. of hours per week: 4			
Exam duration: 6 Hrs	Maximum Marks: 100 (External:50 Internal:50)	Total No. of contact hours: 60	

#### A) Practicals for Endocrinology

- 1. Histological study of various vertebrate endocrine glands using permanent slides / Photographs (Mammals, Crab and Insects).
- 2. Histology of various vertebrate endocrine glands (Microtomy).
- 3. Morphological and histological study of various insect neuroendocrine structures.
- 4. Effect of toxicants on histoarchitecture of various endocrine glands.
- 5. Study of various endocrine disorders using photographs.
- 6. Study of effect of exogenous growth hormone on the growth of fish.
- 7. Study of vaginal smear during oestrous cycle of Rat.

#### B) Practicals for Fishery-I Fresh Water Fish Culture

- Experiments on Water Analysis: Estimation Dissolved Oxygen, Free Carbon dioxide, Estimation of Dissolved Solids, Chlorides, Carbonate, Bicarbonate, Total Alkalinity, Total hardness, Estimation of Biological Oxygen Demand and Chemical Oxygen Demand.
- 2. Estimation of Primary productivity of any local pond, river, lake or reservoir.
- 3. Plankton Analysis: Collection, preservation and estimation of planktons, Quantitative analysis- Enumeration of Zooplanktons by i) drop count method ii) Sedgwick Rafter Cell method, Determination of population density, abundance and dominance of the species. Preparation of Diversity indices
- 4. Collection, identification and classification of locally available fishes
- 5. Collection and Identification of carp spawn and fry.
- 6. Collection and identification of common aquatic insects.
- 7. Collection and identification of common weeds.
- 8. Permanent micro preparation of different kinds of parasites in fishes.
- 9. Preparation of models and designing of cages and pens.
- 10. Visit to Fish farm. Submission of a detailed report on Fish Farm Visit)

#### The examinee shall be required to produce at the practical examination the following:

Practical record book duly signed by teacher in-charge and certified by the Head of the Department as a bonafide work of the examinees.

## DISTRIBUTION OF PRACTICAL MARKS

#### **External Practical Marks**

Experiments from Section A) Endocrinology	
1. Histological Preparation/Experiment	12 Marks
2. Identification, Labeling and Comment on	
Spots / Photographs of endocrine disorders	08 Marks
Experiments from Section B) Entomology-1 Insect classificati	on, morphology and physiol
3. Major Experiments (Water/ Plankton Analysis)	12 Marks
4. Spotting (1 feed ingredient, 1 parasite, 1 aquatic insect, 1	aquatic weed)08 Marks
5. Viva Voce	
TOTAL	50 Marks
nternal Practical Marks	
nternal Practical Marks 1. Certified Practical Record	: 20 Marks
	: 20 Marks : 10 Marks
1. Certified Practical Record	: 10 Marks
<ol> <li>Certified Practical Record</li> <li>Submission of Stained Permanent sides</li> </ol>	: 10 Marks
<ol> <li>Certified Practical Record</li> <li>Submission of Stained Permanent sides</li> <li>Submission of Fish farm Reports/ Model of cages/pens</li> </ol>	: 10 Marks : 05 Marks

Course: DSE III Animal Physiology-I Th- Major Elective			
Subject Code: 3 ZOO 4 No. of Credits: 3 No. of hours per week: 3			
Exam duration: 3 Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 45	

# CO's: 3 ZOO 4

- 1: Animal physiology gives the knowledge of biological processes through the investigation of physiological processes.
- 2: It enables to understand the chemical and molecular processes that occur in and between cells.
- 3: It also provides knowledge about the theoretical processes related to hormonal action.
- 4: Trains the students to perform laboratory exercises in Animal physiology that is applicable to Pathology laboratory, medicine, forensics and pharmaceutical industry.

Units	Syllabus Content	Hours
	Physiology of Digestion:	
Unit I	1.1 Digestion, Absorption, Utilization of Protein and Carbohydrates.	
	1.2 Digestion, Absorption, Utilization of lipid.	
	1.3 Histophysiologies of gastric gland.	8
	1.4 Gastrointestinal Function—Peristalsis, Nervous Control	
	1.5 Gastrointestinal peptides	
	Physiology of Respiration:	
	2.1 Physiology of Respiration: Anatomical and physiological organization of	
	Respiratory system.	
Unit II	2.2 Mechanism of respiration.	7
	2.3 Transport of gases by blood.	
	2.4 Oxygen dissociation curve, CO <sub>2</sub> dissociation curve.	
	2.5 Respiratory center and Neuro Hormonal and Chemical regulation of respiration.	
	Physiology of Heart:	
	3.1 Anatomy and histology of mammalian heart	
	3.2 Structure & function of Myogenic and neurogenic heart.	
Unit III	3.3 Cardiac cycle, Cardiac sound.	7
	3.4 Pulmonary circulation.	
	3.5 Cardiac arrest.	
	Physiology of Circulation:	
	4.1 Composition of Blood.	
	4.2 Regulation of heart beat and blood pressure.	
Unit IV	4.3 Origin and conduction of cardiac impulse.	8
	4.4. Myocardial infarction and cardiomyopathy.	
	4.5. Hormonal control on circulation.	
	Physiology of Excretion:	
	5.1 Urine formation, Ultra filtration, Reabsorption, and Secretion.	
	5.2 Significance of Henley's loop in production of hyper osmotic urine.	
Unit V	5.3 Function of aldosterone, antidiuretic hormone.	8
	5.4 Rennin-Angiotensin system in renal physiology.	
	5.5 Role of kidney in pH regulation and water salt regulation.	

	Lymphatic System:	
	6.1 Lymph-Composition and Formation	
Unit VI	6.2 Functions of lymph.	7
	6.3 Structure and functions of lymph nodes.	
	6.4 Lymphedema.	

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- 2. Bell Davidson: Text book of physiology and Biochemistry
- 3. Bolander F.F.:: Molecular endocrinology
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- 5. Cole S. W.: The practical physiological chemistry.
- 6. Cooper: Poisoning by drugs and chemicals.
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- 9. Garden M.S.: Animal physiology principal and Adaptations.
- 10. Hara & Oserburg; An introduction to crimminalistie.
- 11. Hill R.W.: Comparative physiology of animals
- 12. Hoar W.S.: General and comparative physiology.
- 13. Houssa: Human physiology (McGraw Hill Books Company)
- 14. Hunter& Bornford: Hutchisons Clinical methods
- 15. Hynes: The Biology of polluted water.
- 16. Jacobs M. B.: The analytic toxicology of inorganic poison
- 17. Keil J.B., SamsonWrights a, : Applied Physiology
- 18. Heil E. Joets N.: Physiology (Oxford Uni press ) (1982)
- 19. Klein L: River pollution, causes& effects
- 20. Madhu Raj: Environmental Management of toxic and hazardous chemicals
- 21. Mill peter J.: Comparative neurobiology (Ed Hrbord London
- 22. Modi N.J.: Text Book of toxicology
- 23. Mitchell P.H.: Text Book of General physiology.
- 24. Norman A.W.: Hormones.
- 25. Odum: Fundamental of ecology.
- 26. Osterbong: The crime laboratory
- 27. Philips G.: Environmental physiology.
- 28. Prosser C.L.: Comparative animal physiology.
- 29. Ramkumar: Environmental Biodegradation.

Course: Pr- Major Laboratory 6			
(DSC III.3 Endocrinology + DSE III Animal Physiology-I)			
Subject Code: 3 ZOO 6 No. of Credits: 2 No. of hours per week: 4			
Exam duration: 6 Hrs	Maximum Marks: 100 (External:50 Internal:50)	Total No. of contact hours: 60	

#### A) Practicals for Endocrinology

- 1. Histological study of various vertebrate endocrine glands using permanent slides / Photographs (Mammals, Crab and Insects).
- 2. Histology of various vertebrate endocrine glands (Microtomy).
- 3. Morphological and histological study of various insect neuroendocrine structures.
- 4. Effect of toxicants on histoarchitecture of various endocrine glands.
- 5. Study of various endocrine disorders using photographs.
- 6. Study of effect of exogenous growth hormone on the growth of fish.
- 7. Study of vaginal smear during oestrous cycle of Rat.

## B) Practicals for Animal Physiology-I

- 1. Simple muscle curve Effects of temperature and calcium.
- 2. Estimation of serum creatinine.
- 3. Estimation of serum urea.
- 4. Qualitative analysis of urea
- 5. Quantities estimation of calcium, phosphorus sodium and potassium.
- 6. Separation and identification of amino acids by paper and thin layer chromatography-
- 7. Separation of proteins by paper or SDS-PAGE electrophoresis.
- 8. Determination of Erythrocyte sedimentation rate (E.S.R.)
- 9. Determination of pack cell volume (P.C.V).
- 10. Determination of mean corpuscular volume (M.V.C.)
- 11. Detection of blood by haemin crystals test.
- 12. Estimation of protein in blood.
- 13. Estimation of glucose in given sample.
- 14. Estimation of cholesterol in blood.

# The examinee shall be required to produce at the practical examination the following:

Practical record book duly signed by teacher in-charge and certified by the Head of the Department as a bonafide work of the examinees.

## DISTRIBUTION OF PRACTICAL MARKS

#### **External Practical Marks Experiments from Section A) Endocrinology** 2. Identification, Labeling and Comment on Experiments from Section B) Entomology-1 Insect classification, morphology and physiology **TOTAL** 50 Marks **Internal Practical Marks** : 20 marks 1. Certified Practical Record : 10 marks 2. Submission of Stained Permanent sides 3. Survey and collection of Data of Blood Parameters : 10 marks : 10 marks 4. Student Performance TOTAL 50 Marks

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Course: DSE III Molecular Biology-1 Th- Major Elective-I			
Subject Code: 3 ZOO 4 No. of Credits: 3 No. of hours per week: 3			
Exam duration: 3 Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 45	

# CO's: 3 ZOO 4

- 1: Molecular Biology gives the knowledge of biological processes through the investigation of molecular mechanisms.
- 2: It enables to understand the chemical and molecular processes that occur in and between cells.
- 3: Trains the students to perform laboratory exercises in Molecular Biology that is applicable to medicine, forensics and pharmaceutical industry.

Unit	Contents	Hours
	DNA replication:	
	1.1 Enzymes of DNA replication.	
<b>T</b> T • 4 <b>T</b>	1.2 Mechanism of prokaryotic DNA replication.	0=
Unit-I	1.3 Mechanism of eukaryotic DNA replication.	07
	1.4 Replication of telomeres.	
	1.5 Assembly of newly replicated DNA into nucleosomes.	
	Transcription and its Regulation:	
	2.1 Regulatory elements.	
	2.2 Mechanism of prokaryotic and eukaryotic transcription.	
Unit-II	2.3 Transcription regulation in eukaryotes.	08
	2.4 Transcriptional and post-transcriptional gene silencing.	
	2.5 Transcription regulation in prokaryotes: Lac and Trp operon.	
	3.1 Co -and Post-transcriptional modifications in mRNA, Nuclear export of	
	mRNA, mRNA stability.	
	Translation (Protein synthesis):	
Unit-III	3.2 Genetic code.	08
	3.3 Protein synthesis in prokaryotes and eukaryotes.	
	3.4 Regulation of translation.	
	3.5 Co- and post-traslational modifications of proteins.	
	DNA Markers:	
	4.1 Restriction fragment length polymorphism (RFLP).	
	4.2 Random amplified polymorphic DNA (RAPD).	
	4.3 Amplified fragment length polymorphism (AFLP).	
Unit-IV	4.4 Single-nucleotide polymorphism (SNP).	07
	4.5 <b>Hybridization Techniques:</b> Preparation of radioactive and nonradioactive	
	probes, Southern blotting, Northern blotting, Western blotting,	
	Fluorescence in situ hybridization (FISH).	
	Recombinant DNA Technology (RDT):	
	5.1 Enzymes used in RDT.	
	5.2 Vectors used in RDT: Plasmids, Bacteriophages, Phagemids, Cosmids,	
<b>Unit-V</b>	BACs, PACs, YACs.	07
	5.3 Procedure of RDT.	
	5.4 Applications of RDT.	
	5.5 Construction and screening of genomic and cDNA library.	

	Sequencing and Other Techniques:	
	6.1 <b>DNA Sequencing Techniques:</b> Sanger's dideoxy method,	
	Illumina sequencing, Ion torrent sequencing, Nanopore sequencing,	
	and Single-molecule real-time (SMRT) sequencing.	
Unit-VI	6.2 Protein sequencing by Sanger's method.	08
	6.3 <b>DNA Fingerprinting:</b> Principle, procedure and applications.	
	6.4 Polymerase Chain Reaction: Methodology, modifications, applications,	
	advantages and limitations.	
	6.5 RNA Interference (RNAi): MicroRNA and SiRNAs.	

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- 2. Brown, T. A. Genomes 4. Garland Science.
- 3. Clark David, Pazdernik N. J., and M. R. McGehee. Molecular Biology, 3/Ed. Academic Press.
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- 6. Glick Bernard. **Medical Biotechnology.** ASM Press, Washington DC.
- 7. Karp Gerald. Cell and Molecular Biology, 8/Ed. John Wiley and Sons, Inc.
- 8. Krebs Jocelyn E. Lewin's Genes XII. Jones & Batlett Learning.
- 9. Lodish H. et al. Molecular Cell Biology, 9/Ed. W. H. Freeman and Company.
- 10. Reece Richard J. Analysis of Genes and Genomes. John Wiley & Sons, Ltd.
- 11. Schleif Robert. Genetics and Molecular Biology, 2/Ed. The Johns Hopkins University Press.
- Walker John and R. Raply (Editors). Molecular Biology and Biotechnology, 5/Ed. RSC Publishing.
- 13. Wink M. (Editor). **An Introduction to Molecular Biotechnology**. Wiley-VCH Verlag GmbH and Co.

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Ī	Course: Pr- Major Laboratory 6				
	(DSC III.3 Endocrinology + DSE III Molecular Biology-1)				
Subject Code: 3 ZOO 6		No. of Credits: 2	No. of hours per week: 4		
	Exam duration: 6 Hrs	Maximum Marks: 100	Total No. of contact hours: 60		
	Exam duration, 0 mis	(External:50 Internal:50)	Total 140. of contact flours. of		

### A) Practicals for Endocrinology

- 1. Histological study of various vertebrate endocrine glands using permanent slides / photographs (Mammals, Crab and Insects).
- 2. Histological preparation of various vertebrate endocrine glands (microtomy).
- 3. Morphological and histological study of various insect neuroendocrine structures.
- 4. Effect of toxicants on histoarchitecture of various endocrine glands.
- 5. Study of various endocrine disorders using photographs.
- 6. Study of effect of exogenous growth hormone on the growth of fish.
- 7. Study of vaginal smear during oestrous cycle of Rat.

#### B) Practicals for Molecular Biology-1

- 1. DNA fingerprinting.
- 2. Extraction of DNA from bacteria.
- 3. Extraction of DNA from yeast.
- 4. Extraction of DNA from animal tissue.
- 5. Extraction of DNA from whole blood.
- 6. Determination of molecular size of DNA.
- 7. Restriction digestion.
- 8. Preparation and demonstration of plasmid by gel electrophoresis.
- 9. Isolation and cleaning of DNA fragment of interest from the agarose gel.
- 10. Electrophoretic separation and determination of molecular weight of protein by SDS-PAGE.

### The examinee shall be required to produce at the practical examination the following:

Practical record book duly signed by teacher in-charge and certified by the Head of the Department as a bonafide work of the examinees.

#### DISTRIBUTION OF PRACTICAL MARKS

#### **External Practical Marks Experiments from Section A) Endocrinology** 2. Identification, Labeling and Comment on **Experiments from Section B) Molecular Biology-1** TOTAL 50 Marks **Internal Practical Marks** 1. Certified Practical Record : 20 marks 2. Submission of Stained Permanent sides : 10 marks 3. Submission of Photographs of Stained Gel : 10 marks 4. Student Performance : 10 marks **TOTAL** 50 Marks

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# M. Sc. Zoology (NEPv23) Semester-IV

Course: DSC I.4 Biochemistry Th- Major		
Subject Code: 4 ZOO 1 No. of Credits: 4 No. of hours per week		No. of hours per week: 4
Exam duration: 3 Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 60

# CO's: 4 ZOO 1

- 1: Fundamental properties of elements, their role in formation of biomolecules and in chemical reactions within living organisms.
- 2: Understanding of concepts of aids, bases, indicators, pKa values, etc. Acquiring skill to determine pKa value of amino acids.
- 3: physical and chemical properties of molecules as a linkage of biochemistry.
- 4: Illustrate the metabolism of carbohydrates through various anabolic and catabolic pathways.
- 5: It trains the students to carry out laboratory exercises in biochemistry and biochemical investigations.

Units	Syllabus Content	Hours
	Chemical Foundations of Biochemistry:	
	1.1 pH, pK, acids, bases, buffers, free energy and isomerization.	
	1.2 Entropy, enthalpy and free energy.	
Unit I	1.3 Physical properties of water.	10
	1.4 Bonds and forces stabilizing biomolecules: Covalent bond, Van-der-Waals	
	Electrostatics, hydrogen bonding and hydrophobic interactions.	
	1.5 Biologically important monosaccharides, disaccharides and polysaccharides.	
	Amino acids and Proteins:	
	2.1 Classification and structure of standard amino acids.	
Unit II	2.2 Transamination and deamination.	10
	2.3 Protein structure & folding, Ramachandran plot.	
	2.4 Conjugated proteins: structure and function.	
	Nucleic Acids:	
	3.1 Structure of DNA	
	3.2 Structural polymorphism of DNA (A, B and Z-DNA).	10
Unit III	3.3 Triplex and quadruplex DNA.	
	3.4 Structure and functions of mRNA, rRNA and tRNA.	
	3.5 Circular DNA.	
	Amino Acid and Nucleotide Metabolism:	
	4.1 Ornithine cycle.	
Unit IV	4.2 Biosynthesis of nutritionally non-essential amino acids.	10
	4.3 <i>De novo</i> and salvage pathways of nucleotide biosynthesis.	
	4.4 Degradation of nucleotides.	
	Carbohydrate metabolism:	
	5.1 Glycolysis, its regulation and energetics.	
	5.2 TCA cycle & its regulation.	
<b>T</b> • • <b>T</b>	5.3 Gluconeogenesis and its regulation.	10
Unit V	5.4 Glycogenesis, Glycogenolysis, coordinated regulation.	10
	5.5 Pentose phosphate pathway.	
	5.6 Electron transport complexes, electron transport, oxidative phosphorylation,	
	Energetics of electron transfer.	

	Lipid Metabolism:	
	6.1 Biosynthesis of fatty acids, triglycerides, phospholipids and cholesterol.	
	6.2 β-oxidation of saturated, monounsaturated and polyunsaturated fatty acids.	
Unit VI	$6.3~\alpha$ - and $\omega$ -oxidation of fatty acids.	10
	6.4 Coordinated regulation of fatty acid synthesis and breakdown.	
	6.5 Obesity and the regulation of body mass.	
	6.6 Biosynthesis of ketone bodies.	
	6.6 Biosynthesis of ketone bodies.	

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- 2. Baynes, John W Baynes et al. Medical Biochemistry, 4/Ed. Elsevier Saunders.
- 3. Berg, Jeremy M. et al. Biochemistry, 8/Ed. W. H. Freeman and Company.
- **4.** Campbell, Mary K. et al. **Biochemistry**, 8/Ed. Cengage Learning.
- **5.** Chatterjea, M. N. *et al.* **Textbook of Medical Biochemistry**, 8/Ed. Jaypee Brothers Medical
- **6.** Devlin, T. M. (Editior). **Textbook of Biochemistry**, 7/Ed. John Wiley & Sons, Inc.
- 7. Garrett, Reginald H. Biochemistry, 6/Ed. Cengage Learning.
- **8.** Mathews, Christopher K. **Biochemistry**, 4/Ed. Pearson.
- 9. Moran, Laurence A. et al. Principles of Biochemistry, 5/Ed. Pearson.
- 10. Nelson, D. L. et al. Lehninger Principles of Biochemistry, 8/Ed. Macmillan Learning.
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- 12. Pratt, Charlotte W. Essential Biochemistry, 4/Ed. John Wiley and Sons, Inc.
- 13. Puri Dinesh. Textbook of Medical Biochemistry, 3/Ed. Elsevier.
- 14. Rodwell, Victor W. Harper's Illustrated Biochemistry, 31/Ed. McGraw Hill Education.
- 15. Satyanarayana, U. Biochemistry. Books and Allied (P) Ltd.
- 16. Vasudevan, D. M. et al. Textbook of Biochemistry, 6/Ed. Jaypee Brothers Medical Publishers.
- 17. Voet, D. et al. Biochemistry, 4/Ed. John Wiley and Sons, Inc.
- 18. Voet, D. et al. Fundamentals of Biochemistry, 5/Ed. John Wiley and Sons, Inc.
- 19. Zubay Geoffrey L. et al. Principles of Biochemistr. Wm. C. Brown Publishers.

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Course: DSC II.4 Enzymology and Biostatistics Th- Major			
Subject Code: 4 ZOO 2 No. of Credits: 4 No. of hours per week		No. of hours per week: 4	
Exam duration: 3 Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 60	

# **COs: 4 ZOO 2**

- 1. Enzymology enables to understand the role and activities of various enzymes functioning in the body.
- 2. It also gives some idea about clinical and pharmaceutical applications of enzymes.
- 3. It trains the students to carry out laboratory exercises related to enzyme activity and estimations of enzymes.
- 4. Biostatistics trains the students in handling and analyzing the biological clinical data.

Enzymes-Structure, Classification and Kinetics:  1.1 Introduction and classification of enzymes.  1.2 RNA World Hypothesis  1.3 Features of enzyme's active site.  1.4 Mechanism of enzyme action (Chymotrypsin).  1.5 Kinetics of single substrate and bisubstrate enzyme catalyzed reactions.  1.6 Effect of pH and temperature on enzymes  Enzymes-Categories & Regulation:  2.1 Enzyme Activators and Inhibitors.  2.2 Isozymes, ribozymes and abzymes.  2.3 Allosteric enzymes.  2.4 Cooperativity in enzyme catalysis.  2.5 Zymogen activation.  2.6 Cowalent modification.  Enzymes-Functional Diversity:  3.1 Mechanism of coenzyme action (NAD, FAD, TPP, Pyridoxal phosphate, biotin)  3.2 Enzymes involved in free radical removal.  3.3 Enzymes involved in energy production (DNA replication, DNA repair and transcription).  3.5 Enzymes involved in energy production (Glycolysis and TCA cycle).  Enzyme Applications:  4.1 Applications of Immobilized enzymes.  4.2 Industrial uses of enzymes: Use of lactase in dairy industry  4.4 Enzymes and modern medicine.  4.5 Use of enzymes in fermentation process,  4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics:  5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram).  5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).  5.3 Measurement of central tendency (Mean, median, mode)	Unit	Contents	Hours
Unit-II  1.2 RNA World Hypothesis  1.3 Features of enzyme's active site.  1.4 Mechanism of enzyme action (Chymotrypsin).  1.5 Kinetics of single substrate and bisubstrate enzyme catalyzed reactions.  1.6 Effect of pH and temperature on enzymes  Enzymes-Categories & Regulation:  2.1 Enzyme Activators and Inhibitors.  2.2 Isozymes, ribozymes and abzymes.  2.3 Allosteric enzymes.  2.4 Cooperativity in enzyme catalysis.  2.5 Zymogen activation.  Enzymes-Functional Diversity:  3.1 Mechanism of coenzyme action (NAD, FAD, TPP, Pyridoxal phosphate, biotin)  3.2 Enzymes involved in free radical removal.  3.3 Enzymes involved in ell signaling.  3.4 Enzymes involved in ell signaling.  3.4 Enzymes involved in energy production (Glycolysis and TCA cycle).  Enzyme Applications:  4.1 Applications of Immobilized enzymes.  4.2 Industrial uses of enzymes: Use of lactase in dairy industry  4.3 Use of proteases in food, leather and detergent industry  4.4 Enzymes and modern medicine.  4.5 Use of enzymes in fermentation process,  4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics:  5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram).  5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		Enzymes-Structure, Classification and Kinetics:	
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Unit-II  2.2 Isozymes, ribozymes and abzymes. 2.3 Allosteric enzymes. 2.4 Cooperativity in enzyme catalysis. 2.5 Zymogen activation. 2.6 Covalent modification.  Enzymes-Functional Diversity: 3.1 Mechanism of coenzyme action (NAD, FAD, TPP, Pyridoxal phosphate, biotin) 3.2 Enzymes involved in free radical removal. 3.3 Enzymes involved in cell signaling. 3.4 Enzymes involved in nucleic acid metabolism (DNA replication, DNA repair and transcription). 3.5 Enzymes involved in energy production (Glycolysis and TCA cycle).  Enzyme Applications: 4.1 Applications of Immobilized enzymes. 4.2 Industrial uses of enzymes: Use of lactase in dairy industry 4.3 Use of proteases in food, leather and detergent industry 4.4 Enzymes and modern medicine. 4.5 Use of enzymes in fermentation process, 4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics: 5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram). 5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		Enzymes-Categories & Regulation:	
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2.5 Zymogen activation. 2.6 Covalent modification.  Enzymes-Functional Diversity: 3.1 Mechanism of coenzyme action (NAD, FAD, TPP, Pyridoxal phosphate, biotin) 3.2 Enzymes involved in free radical removal. 3.3 Enzymes involved in cell signaling. 3.4 Enzymes involved in nucleic acid metabolism (DNA replication, DNA repair and transcription). 3.5 Enzymes involved in energy production (Glycolysis and TCA cycle).  Enzyme Applications: 4.1 Applications of Immobilized enzymes. 4.2 Industrial uses of enzymes: Use of lactase in dairy industry 4.3 Use of proteases in food, leather and detergent industry 4.4 Enzymes and modern medicine. 4.5 Use of enzymes in fermentation process, 4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics: 5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram). 5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).	Unit-II	2.3 Allosteric enzymes.	10
2.6 Covalent modification.  Enzymes-Functional Diversity: 3.1 Mechanism of coenzyme action (NAD, FAD, TPP, Pyridoxal phosphate, biotin) 3.2 Enzymes involved in free radical removal. 3.3 Enzymes involved in cell signaling. 3.4 Enzymes involved in nucleic acid metabolism (DNA replication, DNA repair and transcription). 3.5 Enzymes involved in energy production (Glycolysis and TCA cycle).  Enzyme Applications: 4.1 Applications of Immobilized enzymes. 4.2 Industrial uses of enzymes: Use of lactase in dairy industry 4.3 Use of proteases in food, leather and detergent industry 4.4 Enzymes and modern medicine. 4.5 Use of enzymes in fermentation process, 4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics: 5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram). 5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		2.4 Cooperativity in enzyme catalysis.	
Enzymes-Functional Diversity:  3.1 Mechanism of coenzyme action (NAD, FAD, TPP, Pyridoxal phosphate, biotin)  3.2 Enzymes involved in free radical removal.  3.3 Enzymes involved in cell signaling.  3.4 Enzymes involved in nucleic acid metabolism (DNA replication, DNA repair and transcription).  3.5 Enzymes involved in energy production (Glycolysis and TCA cycle).  Enzyme Applications:  4.1 Applications of Immobilized enzymes.  4.2 Industrial uses of enzymes: Use of lactase in dairy industry  4.3 Use of proteases in food, leather and detergent industry  4.4 Enzymes and modern medicine.  4.5 Use of enzymes in fermentation process,  4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics:  5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram).  5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		2.5 Zymogen activation.	
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Unit-III  3.2 Enzymes involved in free radical removal.  3.3 Enzymes involved in cell signaling.  3.4 Enzymes involved in nucleic acid metabolism (DNA replication, DNA repair and transcription).  3.5 Enzymes involved in energy production (Glycolysis and TCA cycle).  Enzyme Applications:  4.1 Applications of Immobilized enzymes.  4.2 Industrial uses of enzymes: Use of lactase in dairy industry  4.3 Use of proteases in food, leather and detergent industry  4.4 Enzymes and modern medicine.  4.5 Use of enzymes in fermentation process,  4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics:  5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram).  5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		Enzymes-Functional Diversity:	
Unit-III  3.3 Enzymes involved in cell signaling. 3.4 Enzymes involved in nucleic acid metabolism (DNA replication, DNA repair and transcription). 3.5 Enzymes involved in energy production (Glycolysis and TCA cycle).  Enzyme Applications: 4.1 Applications of Immobilized enzymes. 4.2 Industrial uses of enzymes: Use of lactase in dairy industry 4.3 Use of proteases in food, leather and detergent industry 4.4 Enzymes and modern medicine. 4.5 Use of enzymes in fermentation process, 4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics: 5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram). 5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		3.1 Mechanism of coenzyme action (NAD, FAD, TPP, Pyridoxal phosphate, biotin)	
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4.2 Industrial uses of enzymes: Use of lactase in dairy industry  4.3 Use of proteases in food, leather and detergent industry  4.4 Enzymes and modern medicine.  4.5 Use of enzymes in fermentation process,  4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics:  5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram,  Pie diagram).  5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative  Frequency).		Enzyme Applications:	
Unit-IV  4.3 Use of proteases in food, leather and detergent industry  4.4 Enzymes and modern medicine.  4.5 Use of enzymes in fermentation process,  4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics:  5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram).  5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		4.1 Applications of Immobilized enzymes.	
4.4 Enzymes and modern medicine. 4.5 Use of enzymes in fermentation process, 4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics: 5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram). 5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		4.2 Industrial uses of enzymes: Use of lactase in dairy industry	
4.5 Use of enzymes in fermentation process, 4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics: 5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram). 5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).	Unit-IV	4.3 Use of proteases in food, leather and detergent industry	10
4.6 Enzyme Engineering and site directed mutagenesis  Biostatistics:  5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram).  5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		4.4 Enzymes and modern medicine.	
Biostatistics:  5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram).  5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		4.5 Use of enzymes in fermentation process,	
Unit- V  5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram, Pie diagram).  5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		4.6 Enzyme Engineering and site directed mutagenesis	
Unit- V Pie diagram).  5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		Biostatistics:	
Unit- V 5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative Frequency).		5.1 Diagrammatic representation of data (Line graph, Histogram, Bar diagram,	
Frequency).		Pie diagram).	
	Unit- V	5.2 Graphic representation of data (frequency polygon, frequency curve, cumulative	10
5.3 Measurement of central tendency (Mean, median, mode)		Frequency).	
		5.3 Measurement of central tendency (Mean, median, mode)	

	5.4 Standard deviation.	
	5.5 Standard error.	
	5.6 Significance test (Students't'- test) - paired and unpaired.	
	Biostatistics:	
	6.1 Chi square test as a test for goodness of fit.	
	6.2 Analysis of variance (ANOVA).	
	6.3 Correlation analysis: Correlation types and methods to study correlation,	
Unit-VI	Significance test of correlation coefficient.	10
	6.4 Regression analysis: Kinds of regression analysis (regression line,	
	Regression Equations).	
	6.5 Applications of Biostatistics in research.	
		I

- 1. Animal Cell Culture A practical approach, Ed. JohnR.W. Masters. IRL Press.
- 2. Introduction to instrumental analysis, Robert Braun. McGraw Hill International Editions.
- A Biologists Guide to Principles and Techniques of Practical Biochemistry. K. Wilson & K.H. Goulding, ELBS Edn.
- 4. Samuel Delvin, Enzymes, Sarup& Sons, N. Delhi
- 5. Practical biochemistry edited by Walker
- 6. Foundation in microbiology: Talaro
- 7. Microbiology: Pelczar
- 8. Biology of microorganisms: Madigan, Martinko and Parker.
- 9. Biophysical chemistry- Principles and technique: Upadhyay, Nath
- 10. Statistical techniques in Bioassay Z.Govidarajulu (2000): Pub.S.Kargar
- 11. Statistical method in Bioassay Pub D.J.Finney (1971):. Griffin
- Laboratory manual for Biochemistry and Molecular biology, Shivnery Publishers R. N. Vankhede
   S. N. Niwane
- 13. Probit analysis D.J. Finney (1971): 3rd edition Pub. Griffin

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Course: Pr- Major Laboratory 7				
(DSC I.4- Biochemistry + DSC II.4 - Enzymology and Biostatistics )				
Subject Code: 3 ZOO 7	No. of Credits: 2	No. of hours per week: 4		
Exam duration: 6 Hrs	Maximum Marks: 100 (External:50 Internal:50)	Total No. of contact hours: 60		

# A) Practicals for Biochemistry

- 1. Determination of isoelectric pH of amino acids.
- 2. Estimation of amino acids by Ninhydrin method.
- 3. Estimation of serum protein.
- 4. Estimation of plasma /serum glucose.
- 5. Estimation of glycogen from tissue.
- 6. Determination of acid value of fat.
- 7. Estimation of serum cholesterol.
- 8. Estimation of phospholipids.
- 9. Estimation of plasma calcium.
- 10. Estimation of Protein by Biuret method (Qualitative Test).
- 11. Isolation of casein from milk.

### B) Practicals for Enzymology and Biostatistics

- 1. Study effect of pH and temperature on enzyme activity. Ex. Salivary amylase.
- 2. To study the effect of inhibitors on enzyme activity.
- 3. Colorimetric estimation of some respiratory enzymes.
- 4. Estimation of plasma / serum glucose.
- 5. Estimation of glycogen from tissue.
- 6. Estimation of serum cholesterol.
- 7. Estimation of phospholipids.
- 8. Estimation of lactate dehydrogenase.
- 9. Estimation of plasma proteins.
- 10. Estimation of Na ions.
- 11. Estimation of K ions.
- 12. Estimation of Calcium.
- 13. Estimation of ATPase.
- 14. Estimation of SGOT / SGPT.
- 15. Estimation of Acetylcholinesterase.
- 16. Estimation of acid alkaline phosphatase.
- 17. Estimation of catalase.
- 18. Examples from Biostatistics as per theory.
- 19. Drawing Pie, Bar (column) and Line diagrams on computer.

Note: Besides these any other additional experiment relevant to the syllabus.

#### The examinee shall be required to produce at the practical examination the following:

Practical record book duly signed by teacher in-charge and certified by the Head of the Department as a bonafide work of the examinees.

# DISTRIBUTION OF PRACTICAL MARKS

# **External Practical Marks**

<b>Experiments from Section A) Biochemistry</b>	
1. Major Estimation/Experiment	
2. Minor Estimation/Experiment	08 Marks
Experiments from Section B) Enzymology and	d Biostatistics
3. Major Estimation/Experiment	12 Marks
4. Minor Estimation/Experiment	
5. Viva Voce	10 Marks
TOTAL	50 Marks

# **Internal Practical Marks**

1. Certified Practical Record	: 20 Marks
2. Submission of elaborative report	
on five biochemical disorders	: 10 Marks
3. Submission of Pie, Bar and Line diagrams on computer	: 10 Marks
4. Student Performance	: 10 Marks
TOTAL	50 Marks

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Course: DSC III.4 General Parasitology Th- Major		
Subject Code: 4 ZOO 3	No. of Credits: 3	No. of hours per week: 3
Exam duration: 3Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 45

# CO's: 4 ZOO 3

- 1. Learn the various types of parasites and hosts.
- 2. Establish the relationship between a parasite and the host and their effects.
- 3. Students can differentiate the parasitic Protozoan, Helminthes, and Nematodes.
- 4. Learn of the geographic distribution, life cycle, pathogenicity control and treatment of Protozoan, Helminthes, Nematode parasites.
- 5. Learn the different vectors of Arthropods.

Unit	Contents	Hours
	Concept of Parasitology	
	1.1 Basic concept of Parasitism.	
	1.2 Symbiosis, phoresis, commensalisms, mutualism, parasitoids.	
Unit-I	1.3 Types of Parasites and hosts.	7
	1.4 Parasitic transmission.	
	1.5 Parasitic adaptations.	
	1.6 Advantages and disadvantages in parasitic life.	
	Protozoan Parasitology:	
	2.1 Characteristics of protozoa parasites. Protozoan parasitic diseases ( <i>Giardiasis</i> ,	
	toxoplasmosis, amoebiasis, African trypanosomiasis)	
Unit-II	2.2 Parasitic protozoa and interaction with host.	8
	2.3 Life cycle and diseases caused by Plasmodium sp.	
	2.4 Vectors of protozoan parasites.	
	2.5 Control strategies.	
	Helminth Parasitology:	
	3.1 General characters of Phylum: Platyhelminthes, Nemathelminthes and	
	Acanthocephala	
	3.2 Classification of helminth parasites.	
Unit-III	3.3 life cycle, transmission, pathogenesis, symptoms, epidemiology, diagnosis and	7
	General control measures of liver fluke Fasciola spp.	
	3.4 life cycle, transmission, pathogenesis, symptoms, epidemiology, diagnosis and	
	General control measures of blood flukes Schistosoma nasale spp.	
	3.5 Larval form of Helminthes.	
	Nematode parasitology:	
	4.1 Classification, general account of nematodes.	
	4.2 Mode of transmission and pathogenicity	
Unit-IV	4.3 Morphology, Life-cycle, Treatment and Prophylaxis of-Wuchereria Sp,	8
	4.4 Ancylostoma Sp	
	4.5 Dracunculus Sp	
	Parasitic Zoonoses:	
<b>.</b>	5.1 Introduction of Zoonosis.	
Unit- V	5.2 Nature and epidemiology of zoonotic viral diseases (Rabies, dengue,	8
	Japanese encephalitis).	

5.3 Bacterial diseases (Brucellosis, Plague).		
5.4 Protozoan diseases (Toxoplasmosis, Trypanosomiasis, Leishmaniasis and		
Babesiosis).		
5.5 Prevention and control of parasitic zoonoses.		
Vector Biology:		
6.1 Vectors and its importance in transmission of parasites.		
6.2 Vector and transmission of diseases- mosquito, tick, lice, fleas.	_	
6.3 Host-vector relationship.	7	
6.4 Mechanical and biological vector.		
6.5 Vector control and prevention of vector borne disease transmission.		
	<ul> <li>5.4 Protozoan diseases (Toxoplasmosis, Trypanosomiasis, Leishmaniasis and Babesiosis).</li> <li>5.5 Prevention and control of parasitic zoonoses.</li> <li>Vector Biology:</li> <li>6.1 Vectors and its importance in transmission of parasites.</li> <li>6.2 Vector and transmission of diseases- mosquito, tick, lice, fleas.</li> <li>6.3 Host-vector relationship.</li> <li>6.4 Mechanical and biological vector.</li> </ul>	

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- 13. Parija S.C. Review of Parasitic Zoonoses. A.I.T.B.S.Publishers and Distributors, Delhi.
- 14. Park K. (2003). Text book of Preventive and Social Medicine. Banarsidas Bhanot Publishers Jabalpur, India.
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Course: DSE IV Entomology-II Th-Major Elective II				
Industrial Entomology and Insect Pest Management				
Subject Code: 4 ZOO 4	No. of Credits: 3	No. of hours per week: 3		
Exam duration: 3Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 45		

### CO's: 4 ZOO 4

- 1: Students are trained in life history and ecology of insect pests and predators/ parasites as well as the basic principles and strategies of their management.
- 2: The knowledge acquired and skill developed in the field of entomology
- 3: Understand the role of insect pathogenic nematodes, viruses, bacteria, fungi, protozoa etc., their mode of action.
- 4: Acquire an understanding about chemical insecticides.
- 5. Develop skill related with sericulture, apiculture and lac culture.

Units	Syllabus Content	Hours
Uillts	Mulberry sericulture:	Hours
Unit I	1.1 Cultivation of food plants.	
	1.2 Rearing of silkworms.	7
	1.3 Harvesting and processing of cocoons	
	1.4 Diseases of <i>Bombyx mori</i> .	
	1.5 Predators and parasitoids of silkworm and their management.	
	Apiculture:	
	2.1 Organization of bee colony.	
	2.2 Dance language of honeybees.	
Unit II	2.3 Diseases of honeybees.	8
	2.4 Beekeeping methods: Equipment and tools	
	2.5 Apiary management.	
	Lac culture:	
	3.1 Lac insect and its life history.	7
	3.2 Host plant management.	
Unit III	3.3 Strains of lac insects, Propagation of lac insects.	
	3.4 Lac crop management.	
	3.5 Natural enemies of lac insects and their management.	
	3.6 Lac extraction and lac products.	
	Nature and extent of damage, seasonal abundance of followings:	
	4.1 Insect pests of cereals and millets.	
	4.2 Insect pests of pulses, tobacco, oilseeds.	8
Unit IV	4.3 Insect pests of fiber crops, forages, sugarcane.	
	4.4 Insect pests of Fruit Crops- mango, guava, banana citrus.	
	4.5 Major Insect pests of tomato, Brinjal, okra.	
	4.6 Major Insect pests of stored grain.	
	Biological control of pest:	
	5.1 important groups of Parasitoids, predators and pathogens for pest control.	
TI!4 T7	5.2 Augmentation and conservation.	7
Unit V	5.3 Host seeking behavior of predatory and parasitic groups of insects.	
	5.4 Role of insect pathogenic nematodes, viruses, bacteria, fungi, protozoa etc	
	5.5 Biological control of weeds using insects.	

	Chemical control of Insect pest:	
Unit VI	6.1 Pesticide use and pesticide industry in India.	
	6.2 Classification of insecticides and acaricides based on mode of entry, mode	
	of action and chemical nature.	
	6.3 Structure and mode of action of organochlorines, organophosphates,	8
	Carbamates, pyrethroids.	
	6.4 Action of insecticides- synergism, potentiation and antagonism	
	6.5 Pest resistance to insecticides; mechanisms and types of resistance	
	6.6 Insecticide resistance management and pest resurgence.	

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- Ganga G. 2003. Comprehensive Sericulture. Vol. II. Silkworm Rearing and Silk Reeling. Oxford & IBH, New Delhi.
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- Dhaliwal GS & Arora R. 2003. Integrated Pest Management Concepts and Approaches. Kalyani Publ., New Delhi.
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Course: Pr- Major Laboratory 8 (DSC I.4- General Parasitology + DSE IV Entomology-II- Industrial Entomology and		
Insect Pest Management )		
Subject Code: 3 ZOO 8	No. of hours per week: 4	
Exam duration: 6 Hrs	Maximum Marks: 100 (External:50 Internal:50)	Total No. of contact hours: 60

- Slides / museum specimens of selected parasites of representative groups of protozoans and Helminths (Plasmodium, Trypanosoma, Leishmenia, Ascaris (male & female), Hookworm, Ancylostoma sp., Wucheria sp..
- 2. Study of life cycles and morphology of Vectors: Anopheles, Culex and, Aedes species (Adults, eggs, larvae and pupae), house fly, cockroach, bed bug.
- 3. Ticks and mites: Argus, Sarcoptes, Psoroptes, Hemaphysalis spinigera
- 4. Collection of ticks and mites from domestic animals.
- 5. Histological preparation of parasites in tissues.
- 6. Preparation of blood smear for protozoan / nematode parasites.

#### B) Practicals for Entomology-II- Industrial Entomology and Insect Pest Management

- 1. Identification of immature insects to orders and families, in endopterygote orders viz., Diptera, Lepidoptera, Hymenoptera and Coleoptera using key.
- 2. Identification of honey bee species, bee castes and special adaptations.
- 3. Visit to bee nursery and commercial apiaries.
- 4. Visit to Silkworm rearing and management center.
- 5. Field collection of parasitoids and predators.
- 6. Identification of common natural enemies of crop pests (parasitoids, predators, microbes) and weed killers.
- 7. Control of silverfishes in the library
- 8. Study of life history of important insect pests and non-insect pest.
- 9. Lac host and crop management technology and processing of lac.

Note: Besides these any other additional experiment relevant to the syllabi depending on resource.

#### The examinee shall be required to produce at the practical examination the following:

Practical record book duly signed by teacher in-charge and certified by the Head of the Department as a bonafide work of the examinees.

# DISTRIBUTION OF PRACTICAL MARKS

#### **External Practical Marks Experiments from Section A) General Parasitology** Experiments from Section B) Entomology-II- Industrial Entomology and Insect Pest Management 3. Identification and classification of insects Pest (photograph/slides) (04 spots).... 08 Marks **TOTAL** 50 Marks **Internal Practical Marks** 1. Certified Practical Record : 20 marks 2. Submission of collection of ticks and mites : 10 marks 3. Submission of case study of successfully Biological control project : 10 marks 4. Student Performance : 10 marks

TOTAL 50 Marks

Course: DSE IV Fisheries II Major Elective			
Fish Harvesting, Marketing and Aquarium Management			
Subject Code: 4 ZOO 4 No. of Credits: 3 No. of hours per week: 3			
Exam duration: 3Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 45	

#### **COs**

- 1. Students will get familiar with the various fishing methods.
- 2. Students will gain knowledge about fish spoilage and causative agents.
- 3. Students will know about different seafood packaging materials and methods of packaging and transport.
- 4. Students will acquire knowledge on quality assurance in fish processing
- 5. Students will acquire a skill of aquarium fabrication and maintenance.

Unit	Course Content	Hours
	1.1 Traditional methods of fish harvesting: Traps, Ring seine, Trammel net, Mini	
Unit I	trawls, Gill nets, Hook and line, traps and pots.	
	1.2 Modern methods of fish harvesting: Trawling, Purse seining, Gill net, Hook	
	and line mechanized.	07
Omt 1	1.3 Unconventional fishing methods: Electrofishing, light fishing, Echo sounder	07
	and sonar.	
	1.4 Biological factors in fishing.	
	1.5 Fishing crafts- Mechanized and non-mechanized boats.	
	2.1 Biochemical composition and nutritional value of fish.	
	2.2 Fish decomposition- Post mortem changes and rigor mortis, Causes of	
	spoilage.	
T1 14 TT	2.3 Fish handling methods: Transferring catch from gear to vessel,	00
Unit II	Washing/Sorting, Bleeding/gutting,	08
	2.4 Traditional methods of fish processing: - Icing, Drying, Salting, Smoking,	
	2.5 Fish Products and byproducts: Fish Oil, Fish meal, Fish silage, Fish protein,	
	Fish glue, Fish leather.	
	3.1 Principles and importance of fish preservation.	
	3.2 Fish Preservation techniques: Icing of fish, different types of ice and their	
	manufacture. CSW, RSW.	
T1:4 TTT	3.3 Different types of freezers. Quality changes during frozen storage.	08
Unit III	3.4 Basic refrigeration cycle, Canning: Unit steps in canning and their	Uð
	significance.	
	3.5 Packaging of fish and fishery products:- Packaging materials; basic films and	
	laminates,	
	4.1 Fish Markets and Fish market structure.	
	4.2 Types of market: wholesale, terminal, retail, and fairs. Functions: Selling,	
	transportation, storage, gradation, money transaction.	
Unit IV	4.3 Marketing system: Use flows, physical flows and channel flows.	07
	4.4 Government and Fishermen's Co-operative Societies, integration, marketing	
	efficiency.	
	4.5 Price determination, Institutional Support to fisheries, Crop Insurance.	
Unit V	5.1 Quality Assurance and Export of Fishery Products: Quality control – basic	07
Cint v	concepts, quality and quality control.	U/

	5.2 Sanitation procedures in fish processing plants.	
	5.3 Risk factors in fish bio-toxins, seafood pathogens, endogenous parasites.	
	5.4 Methods of evaluating fish freshness and quality - organoleptic, physical,	
	chemical, microbiological and instrumental methods.	
	5.5 Quality standards in India and major importing countries like USA, Japan and	
	EU.	
	6.1 Aquarium design and Construction: Introduction to aquarium.	
	6.2 Design and construction of home and public aquaria (freshwater and marine),	
	Aquarium accessories	
	6.3 Aquarium Management: Setting up of aquarium – under gravel filter, pebbles,	
Unit VI	plants, drift wood, ornamental objects and selection of fishes.	08
	6.4 Aquarium maintenance and water quality management. Control of snail and	
	algal growth. Handling, care, packing and transportation of fishes	
	6.5 Freshwater Ornamental Fishes: Species of ornamental fishes - their taxonomy	
	and biology- Live bearers, Gold fish and koi,	

#### **Suggested Readings:**

- 1. ADCP (Aquaculture Development and Co-ordination Programme). 1980.
- 2. Badapanda K.C. Fishing Crafts and Gear technology. Narendra Publishing House.
- 3. Balachandran KK. 2001. Post-harvest Technology of Fish and Fish Products. Daya Publ. House.
- 4. Biswas K.P. Advancement in fish, fisheries and technology. Narendra Publishing House.
- 5. Charls L. Cutting, 2002. Fish Processing and Preservation.
- 6. Clucas IJ. 1981. Fish Handling, Preservation and Processing in the Tropics. Parts I, II. FAO.
- 7. Fennema K, Powrie WD & Marth EH. 1973. Low Temperature Preservation of Foods and Living Matter. Marcel Dekker.
- 8. Gopakumar K. (Ed.). 2002. Text Book of Fish Processing Technology. ICAR.
- 9. Govindan T.K. Fish Processing Technology. Oxford & IBH Pub. Co.
- 10. Hall GM. (Ed). 1992. Fish Processing Technology. Blackie.
- 11. Hall GM. (Ed). 2011. Fish Processing –sustainability and new opportunities. Wiley Blackwell.
- 12. Hand book of Fisheries and Aquaculture. 2006. Indian Council of Agricultural Research. New
- 13. Hersom AC & Hulland ED. 1980. Canned Foods. Chemical Publ. Co.
- 14. Hertrampf JW & Pascual FP. 2000. Handbook on Ingredients for Aquaculture Feeds. Kluwer. Houlihan D, Boujard T & Jobling M. 2001. Food Intake in Fish. Blackwell.
- 15. Larousse J & Brown BE. 1997. Food Canning Technology. Wiley VCH.
- 16. Lavens P & Sorgeloos P. 1996. Manual on the Production and Use of LiveFood for Aquaculture. FAO Fisheries Tech. Paper 361,
- 17. M.N. Moorjani, 1998. Fish Processing in India. ICAR, New Delhi,
- 18. Manual on Fish Canning. FAO Fisheries Tech. Paper 285.
- 19. Post harvest technology of freshwater fish. 2009. Central institute of fisheries technology. Cochin.
- 20. Saroj K. Swain, Sarangi N. and Ayyappan S. 2010. Ornamental Fish Farming ICAR
- 21. Sen DP. 2005. Advances in Fish Processing Technology. Allied Publ.
- 22. Thabrow De WV. 1981. Popular Aquarium Plants. Thornbill Press.
- 23. Ujwala Jadhav (2010). Aquaculture Technology and Environment. Publ. PHI Publication.
- 24. Venugopal V. 2006. Seafood Processing. Taylor & Francis.

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Course: Pr- Major Laboratory 8			
(DSC I.4- General Parasitology + DSE IV Fisheries-II- Fish Harvesting, Marketing and			
Aquarium Management )			
Subject Code: 3 ZOO 8 No. of Credits: 2 No. of hours per week:			
Exam duration: 6 Hrs	Maximum Marks: 100 (External:50 Internal:50)	Total No. of contact hours: 60	

- 1. Slides / museum specimens of selected parasites of representative groups of protozoans and helminths (Plasmodium, Trypanosoma, Leishmenia, Ascaris( male & female), Hookworm, Ancylostoma sp., Wucheria sp..
- 2. Study of life cycles and morphology of Vectors: Anopheles, Culex and, Aedes species (Adults, eggs, larvae and pupae), house fly, cockroach, bed bug.
- 3. Ticks and mites: Argus, Sarcoptes, Psoroptes, Hemaphysalis spinigera
- 4. Collection of ticks and mites from domestic animals.
- 5. Histological preparation of parasites in tissues.
- 6. Preparation of blood smear for protozoan / nematode parasites.

# B) Practicals for Fisheries-II- Fish Harvesting, Marketing and Aquarium Management

- 1. Study of local traditional fishing gears.
- 2. Identification of fishing crafts, gears, fishing accessories (floats/sinkers/hook/synthetic and natural fibres, twines, ropes, iron wares).
- 3. Evaluation of freshness of fish
- 4. Determination of condition factor of a fish.
- 5. Quality control of fishes: Crude protein analysis of fish muscle by lowry method.
- 6. Identification of tools and accessories used in aquarium fabrication and maintenance.
- 7. Construction of a glass aquarium.
- 8. Identification of aquarium fishes.
- 9. Identification of aquarium plants
- Visit to fish processing plant: Submission of a detailed report of visit.
- Local fish market Survey and submission of its report

# The examinee shall be required to produce at the practical examination the following:

Practical record book duly signed by teacher in-charge and certified by the Head of the Department as a bonafide work of the examinees.

DISTRIBUTION OF PRACTICAL MAI	RKS
External Practical Marks	
Experiments from Section A) General Parasitology	
1. Identification of spots.	12 Marks
2. Study of life cycle/morphological identification of vector	rs
Experiments from Section B) Fisheries-II- Fish Harvesting, Marketin	ng and Aquarium Management
3. Experiments (Expt. 1 or 2 or 3)	12 Marks
4. Spotting (1 fishing accessory, 1 aquarium tool, 1 Aquari	um fish, 1 aquarium plant) 08 M
5. Viva Voce	
TOTAL	50 Marks
Internal Practical Marks	
1. Certified Practical Record	: 20 marks
2. Submission of collection of ticks and mites	: 10 marks
3. Submission of Reports (Visit to fish processing plant: and	d Local fish market Survey: 05 M
4. Setting of Aquarium	: 05 Marks
5. Student Performance	: 10 marks
TOTAL	50 Marks

Course: DSE IV Animal Physiology – II Th-Major Elective		
Subject Code: 4 ZOO 4	No. of Credits: 3	No. of hours per week: 3
Exam duration: 3Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 45

# CO's: 4 ZOO 4

- 1: Animal physiology gives the knowledge of biological processes through the investigation of physiological processes.
- 2: It enables to understand the chemical and molecular processes that occur in and between cells.
- 3: It also provides knowledge about the theoretical processes related to hormonal action.
- 4: Trains the students to perform laboratory exercises in Animal physiology that is applicable to Pathology laboratory, medicine, forensics and pharmaceutical industry.

Units	Syllabus Content	Hours
	Muscle Physiology:	
	1.1 Ultra structure of skeletal muscle and Sarcotubular system.	
	1.2 Ion distribution.	
	1.3 Types of muscle contraction.	0
Unit I	1.4 Muscle proteins.	8
	1.5 Physical and Chemical Properties skeletal muscles.	
	1.6 Sliding filament theory of muscle contraction.	
	1.7 Role of Ca <sup>++</sup> , Calcium receptors, Calmodulin and calcium pump.	
	Nerve Physiology:	
	2.1 Ultra structure of neuron, Electrical properties of nerve.	
	2.2 Ionic concentration in the cytoplasm (Donnan equilibrium system).	
	2.3 Action potential, Resting potential, Depolarization and Repolarization.	
Unit II	2.4 Local circuit theory, ionic theory, Saltatory and nerve conduction.	7
	2.5 Ultrastructure of synapse, Biosynthesis, storage and release of acetylcholine.	
	2.6 Acetylcholine receptor and role of acetylcholine esterase.	
	2.7 Types of neurotransmitters.	
	Homeostasis Physiology:	
	3.1 Homeostasis Physiology: Water contents and distribution	
	3.2 Composition of ECF (Extra cellular fluid) and ICF (Intracellular fluid)	
Unit III	3.3 Abnormal water and electrolyte metabolism and water intoxication.	7
	3.4 Maintenance of pH.	
	3.5 Components of Homeostatic Control system.	
	3.6 Reflexes, Local Homeostatic Responses.	
	Control of Homeostasis:	
	4.1 Nitrogen excretion among different animal Groups.	
	4.2 Mechanism of calcium and phosphate Homeostasis.	
Unit IV	4.3 Homeostasis of iron maintenance.	8
	4.4 Homeostasis mechanism of fever.	
	4.5 Homeostatic mechanism of minerals.	
	4.6 Homeostasis and antidiuretic hormone	
	Thermoregulation and Osmoregulation:	
	5.1 Adaptation and Acclimatization. Biological Rhythms in body temperature.	
Unit V	5.2 Balance in the Homeostasis of chemicals	8
	5.3 Homeostatic control systems - feedback.	

	5.4 Basic thermoregulatory mechanism in Poikilotherms and endotherms.	
	5.5 Ectothermic adaptations to extreme temperatures.	
	5.6 Osmoregulatory mechanism in stenohaline and euryhaline speciesand in	
	Terrestrial environment.	
	Senses and Receptors:	
	6.1 Mechanoreceptors.	
	6.2 Photo receptors.	
Unit VI	6.3 Thermo receptors.	7
	6.4 Chemoreceptors.	
	6.5 Electroreceptors.	
	6.6 Magneto receptors	

# **Suggested Reading:**

- 1 Baileys: Text book of Histology
- 2 Bell Davidson: Text book of physiology and Biochemistry
- 3 Bolander F.F.: Molecular endocrinology
- 4 Clerk E.E.C. Isolation and identification of Drugs in pharmaceutical of body fluid and post martical Vol.I & II.
- 5 Cole S. W.: The practical physiological chemistry.
- 6 Cooper: Poisoning by drugs and chemicals.
- 7 Eckert, Marsall: Animal physiology mechanism and Adaptations
- 8 Eckert & Ranadak: Animal physiology (CBS) 2nd ED (1978)
- 9 Garden M.S.: Animal physiology principal and Adaptations.
- 10 Hara & Oserburg; An introduction to crimminalistie.
- 11 Hill R.W.: Comparative physiology of animals
- 12 HoarW.S.: General and comparative physiology.
- 13 Houssa: Human physiology (McGraw Hill Books Compny)
- 14 Hunter& Bornford: Hutchisons Clinical methods
- 15 Hynes: The Biology of polluted water.
- 16 Jacobs M. B.: The analytic toxicology of inorganic poison
- 17 Keil J.B., Samson Wrightsa, : Applied Physiology
- 18 Heil E. Joets N.: Physiology (Oxford Uni press ) (1982)
- 19 Klein L: River pollution, causes& effects
- 20 Madhu Raj: Environmental Management of toxic and hazardous chemicals
- 21 Mill peter J.: Comparative neurobiology (EdHrbord London
- 22 Modi N.J.: Text Book of toxicology
- 23 Mitchell P.H.: Text Book of General physiology.

- 24 NormanA.W.: Hormones.
- 25 Odum: Fundamental of ecology.
- 26 Osterbong: The crime laboratory
- 27 Philips G.: Environmental physiology.
- 28 ProsserC.L.: Comparative animal physiology.
- 29 Ramkumar: Environmental Biodegradation.
- 30 Ramkumar: Environmental Chemical hazards.
- 31 Robert & Cosselin:First & emergency treatment and clinical toxicology of commercial product.
- 32 Seinfield J.J.: Air pollution(A,P.)
- 33 Smith ptterson: Text Book of physiology (ELBS) Read & Scratched (1988) llth Ed..
- 34 Sern A.C.: Air pollution (A.P.)
- 35 Stewart& stratman: Toxicology mechanism and analytical methods
- 36 Theils: Clinical Toxicology.
- 37 Tomb: An introduction to invertebrate endocrinology (Academic press)
- 38 West Best &Taylor,s: Physiological Basis of medical practice.
- 39 White R. Steions.: Pesticides in environment Vol.1
- 40 Wilsom J. A.: Principles of animal physiology.
- 41 Wod Dennus W.: Principles of animal physiology.(Ed.Arbod) Lond

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Course: Pr- Major Laboratory 8			
(DSC I.4- General Parasitology + DSE IV Animal Physiology - II)			
Subject Code: 3 ZOO 8 No. of Credits: 2 No. of hours per wee			
Exam duration: 6 Hrs	Maximum Marks: 100 (External:50 Internal:50)	Total No. of contact hours: 60	

- Slides / museum specimens of selected parasites of representative groups of protozoans and Helminths (Plasmodium, Trypanosoma, Leishmenia, Ascaris (male & female), Hookworm, Ancylostoma sp., Wucheria sp..
- 2. Study of life cycles and morphology of Vectors: Anopheles, Culex and, Aedes species (Adults, eggs, larvae and pupae), house fly, cockroach, bed bug.
- 3. Ticks and mites: Argus, Sarcoptes, Psoroptes, Hemaphysalis spinigera
- 4. Collection of ticks and mites from domestic animals.
- 5. Histological preparation of parasites in tissues.
- 6. Preparation of blood smear for protozoan / nematode parasites.

# B) Practicals for Animal Physiology – II

- 1. Properties of saliva. Isolation and identification of rumen microorganisms.
- 2. Estimation of rumen ammonia and blood urea under various physiological conditions.
- 3. Normal and abnormal constituents of urine.
- 4. Microscopic examination of urine.
- 5. Preparation and examination of blood smear to study blood cells.
- 6. Differential leucocytes count.
- 7. Histochemical demonstration of-
  - Carbohydrates,
  - Proteins,
  - Lipids.
  - Nucleic acids,
  - Acid and alkaline phosphatase.
- 8. Separation of proteins by paper and gel electrophores
- 9. Qualitative analysis of urea, ketone bodies and salts

#### The examinee shall be required to produce at the practical examination the following:

Practical record book duly signed by teacher in-charge and certified by the Head of the Department as a bonafide work of the examinees.

#### DISTRIBUTION OF PRACTICAL MARKS

# **External Practical Marks**

# **Experiments from Section A) General Parasitology**

2. Study of life cycle/morphological identification of vector	s 08 Marks
Experiments from Section B) Animal Physiology – II	
3. Major physiology experiments	12 Marks
4. Minor physiology experiments	08 Marks
5. Viva Voce	10 Marks

TOTAL 50 Marks

#### **Internal Practical Marks**

1. Certified Practical Record	: 20 marks
2. Submission of collection of ticks and mites	: 10 marks
3. Survey based on physiology practical	: 10 marks
4. Student Performance	: 10 marks

TOTAL 50 Marks

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Course: DSE IV Molecular Biology-II Th-Major Elective ( Molecular Immunology)		
Subject Code: 4 ZOO 4	No. of Credits: 3	No. of hours per week: 3
Exam duration: 3Hrs	Maximum Marks: 100 (External:60 Internal:40)	Total No. of contact hours: 45

#### CO's: 4 ZOO 4

- 1: Molecular Immunology gives the knowledge of biological defence processes through the investigation of molecular mechanisms.
- 2: It enables to understand the physiological and molecular mechanisms that occur in the body during host defence to parasitic infections.
- 3: It gives an idea about various aspects of vaccines and their development.
- 4: Trains the students to perform laboratory exercises in Molecular Immunology that is applicable to medicine and pharmaceutical industry.

Units	Syllabus Content	Hours
	The Immune System:	
Unit I	1.1 Innate and adaptive immunity.	
	1.2 Cells and lymphoid tissues of the immune system.	
	Antigens and Antibodies:	
	1.3 Factors affecting antigenicity.	
	1.4 Epitopes, Haptens, Adjuvants, Superantigens.	
	1.5 Structural features and biological properties of immunoglobulins,	
	Immunoglobulin variants.	
	1.6 Antibody mediated effecter functions.	
	Complement System:	<u> </u>
	2.1 Pathways of complement activation, Late steps of complement activation.	
	2.2 Receptors for complement proteins, Regulation and functions of complement system.	
	Cytokines:	
	2.3 General properties, Cytokines of hematopoiesis, innate and adaptive immunity.	
Unit II	2.4 Cytokine receptors, Cytokine receptor-mediated signal transduction.	07
	Major Histocompatibility Complex (MHC):	
	2.5 Genomic organization, Diversity of MHC.	
	2.6 Structure of MHC molecules, Processing of protein antigens, Binding of peptides to	
	MHC molecules.	
	Biology of T- and B-lymphocytes:	
	3.1 T-Cell maturation, activation, differentiation, and effector functions.	
	3.2 Structure of TCR and TCR complex, Generation of TCR diversity and T-cell	
Unit III	coreceptors.	08
	3.3 Signal transduction by TCR complex.	
	3.4 B-cell maturation, activation, differentiation.	
	3.5 Structure and signal transduction by BCR complex.	
	Techniques in Immunology:	
	4.1 Enzyme-linked immunosorbent assay (ELISA), Radioimmunoassay (RIA).	
Unit IV	4.2 Hybridoma technology, Applications of monoclonal antibodies.	07
CIMU I V	Immunization:	37
	4.3Active Immunization (Vaccines): Live attenuated vaccines, Inactivated or killed	

	vector vaccines, DNA vaccines.	
	<b>4.4 Passive Immunization:</b> Passive immunization through placental antibody	
	transfer and colostrum,	
	5.1 Autoimmunity: Immunologic tolerance, Pathogenesis of autoimmunity.	
	5.2 Autoimmune Diseases: Organ-specific autoimmune diseases,	
Unit V	Systemic autoimmune diseases.	07
	5.3 Immunodeficiency Disorders: Severe combined immune deficiencies (SCIDs),	07
	Acquired immunodeficiency syndrome (AIDS).	
	5.4 <b>Hypersensitivity Reactions:</b> Type I, II, III and IV.	
	Transplantation Immunology:	
	6.1 Immune responses to allografts, Mechanisms of allograft rejection.	
	6.2 Prevention of allograft rejection.	
IImit X/I	Tumor Immunology:	08
Unit VI	6.3 Tumor antigens.	Uo
	6.4 Immune responses to tumors.	
	6.5 Evasion of immune responses by tumors.	

# **Suggested Reading:**

- 1. Abbas Abul K. et al. Cellular and Molecular Immunology, 6-10/Ed. Elsevier.
- 2. Punt Jenni et al. Kuby Immunology, 6-8/Ed. W. H. Freeman and Company New York.
- 3. Coico Richard *et al.* **Immunology-A Short Course**, 6-7/Ed. Wiley-Blackwell.
- 4. Tizard Ian R. Immunology–An Introduction, 4/Ed. Saunders College Publishing.
- 5. Tizard Ian R. **Veterinary Immunology**, 10/Ed. Elsevier.
- 6. Gangal Sudha et al. Textbook of Basic and Clinical Immunology. Orient Blackswan.
- 7. Male David et al. Immunology, 8/Ed. Mosby Elsevier.
- 8. Peter J. Delves *et al.* **Roitt's Essential Immunology**, 13/Ed. Wiley-Blackwell.
- 9. Rich Robert R. et al. Clinical Immunology-Principles and Practice, 5/Ed. Elsevier.
- 10. Flajnik Martin F. et al. Paul's Fundamental Immunology, 8/Ed. Wolters Kluwer.

Course: Pr- Major Laboratory 8		
(DSC I.4- General Parasitology + DSE IV Molecular Biology-II Molecular Immunology)		
Subject Code: 3 ZOO 8	No. of Credits: 2	No. of hours per week: 4
Exam duration: 6 Hrs	Maximum Marks: 100 (External:50 Internal:50)	Total No. of contact hours: 60

- Slides / museum specimens of selected parasites of representative groups of protozoans and Helminths (Plasmodium, Trypanosoma, Leishmenia, Ascaris (male & female), Hookworm, Ancylostoma sp., Wucheria sp..
- 2. Study of life cycles and morphology of Vectors: Anopheles, Culex and, Aedes species (Adults, eggs, larvae and pupae), house fly, cockroach, bed bug.
- 3. Ticks and mites: Argus, Sarcoptes, Psoroptes, Hemaphysalis spinigera
- 4. Collection of ticks and mites from domestic animals.
- 5. Histological preparation of parasites in tissues.
- 6. Preparation of blood smear for protozoan / nematode parasites.

# B) Practicals for Molecular Biology-II Molecular Immunology

- 1. Blood group identification- A, B, AB, O and Rh.
- 2. Screening of antigen and antibody-Antigen-antibody pattern (Ouchterlony double diffusion).
- 3. Estimation of antigen and antibody content in the samples by Radial Immunodiffusion.
- 4. Counter-current immunoelectrophoresis.
- 5. Dot-ELISA.
- 6. Rocket immunoelectrophoresis.
- 7. Immunological diagnosis of pregnancy.
- 8. Preparation of tissue sections and staining of thymus, spleen, and lymph nodes (Source of tissue: Animal tissues from local recognized slaughter houses/ poultry farms/ fish market etc)

### The examinee shall be required to produce at the practical examination the following:

Practical record book duly signed by teacher in-charge and certified by the Head of the Department as a bonafide work of the examinees.

# DISTRIBUTION OF PRACTICAL MARKS

External Pra	DISTRIBUTION OF PRACTICAL MARKS	3
Experi	ments from Section A) General Parasitology	
1.	Identification of spots.	
2.	Study of life cycle/morphological identification of vectors	08 Marks
Experir	nents from Section B) Molecular Biology-II Molecular Imi	munology
3.	Major Immunology experiments	12 Marks
4.	Minor Immunology experiments	08 Marks
5.	Viva Voce	
	TOTAL	50 Marks
nternal Prac	tical Marks	
1. 0	Certified Practical Record	: 20 Marks
2. 3	Submission of collection of ticks and mites	: 10 Marks
3.	Submission of Stained Permanent Slides	: 10 Marks
4.	Student Performance	: 10 Marks
	TOTAL	50 Marks

# Guidelines for PG Research Project (Phase I and Phase II)

- The Research proposal will be evaluated only at college / department level RAC (Research Advisory Committee) and there should be no need to go to the RRC at the University level.
- The department/ college may put up the list of Research Projects (Titles) on the website of concerned RAC/ College / Department in public domain for transparency and information of stakeholders and concerned people.
- All students shall adhere to the following reference guidelines in order to select, pursue and execute
- Research Project during the third and fourth semester of PG Programme. Further, they should adopt
  the following guidelines for preparing and submitting the Report/Thesis of the Research Project.
- The examiners / evaluators are requested to assess the Research Project Thesis on the basis of the following parameters, as applicable.

#### 1. Research Project Title

Title should be clear and concise with appropriate variables (shows accurately the subject/focus area and scope of study through important "keywords" from the subject

The following parameters can be used to help formulate a suitable research project title:

- 1. The purpose of the research
- 2. The type of the research
- 3. The methods used

The initial aim of a title is to capture the reader's attention and to draw his or her attention to the research problem being investigated.

Title of Research project should have several characteristics as follows

- Avoid using abbreviations.
- Use words that create a positive impression and stimulate reader interest.
- Use current nomenclature from the field of study.
- Identify key variables, both dependent and independent.
- Suggest a relationship between variables which supports the major hypothesis.
- Titles are usually in the form of a phrase, but can also be in the form of a question.
- Use correct grammar and capitalization with all first words and last words capitalized, include in the first word of a subtitle. All nouns, pronouns, verbs, adjectives, and adverbs that appear between the first and last words of the title are also capitalized. Simply put, a student must point out several things: (1) what; (2) how; and ideally, (3) why in a Title of the research project.

# 2. Table of Contents, Chapterization Scheme of the Research Project Thesis, List of Tables, List of Figures

#### 3. Research Project Summary (250 words) followed by Keywords

#### 4. Introduction

Origin of the proposal, Motivation for undertaking research, brief overview explaining the background and importance of the study

#### 5. Statement of the Problem Specifically what the researcher wants to know

#### 6. Purpose/Significance/Importance of the Proposed Research in the context of the current status

Topic is critical in discipline. Aim of the proposed work is Unexplored/underexplored Importance/justification of the proposed work. What the researcher hopes to achieve by conducting the study. As part of the purpose of the study, there should be justification for conducting the project. This

section should exhibit a clear understanding of what makes your study significant and why it should be conducted.

#### 7. Definition of Terms, if applicable -

Clarification of any terminology in the study/research that may not be commonly known; provides a similar interpretation for all readers of the study

#### 8. Delimitations, Limitations, and Assumptions (if applicable)

A brief statement identifying the delimitations, limitations, and assumptions associated with the study/research should be provided.

**Delimitations** – factors that were controlled by the researcher

Limitations – factors that were not under the control of the researcher

**Assumptions** – factors that the researcher assumes were taken into consideration.

#### 9. Critical Review of Non-Patent and Patent Literature till date

The student should provide a breakdown of sub-topics influencing the processes of the research project. Each sub-topic should contain a thorough examination of the literature that influences or is representative of current research on that subtopic. The literature review should collectively support the process and purpose of the study. A theoretical framework as applicable to the field of study may be included here. Include Journal Articles (non-patent literature) and Patents (Indian and International) literature (granted/published), if applicable. For Patent literature, student may use, for example, Google Patents Advanced Search. Student shall only include reported research work published in the Journals, which are listed in UGC CARE List Group-I and UGC CARE List Group-II.

# 10. Theoretical Framework, Research Questions, Hypothesis, elements (As applicable to the Research)

Hypothesis must be Clear, corresponding to objectives and testable. Derived from residue of Review of Literature

#### 11. Problem Definition (Formulation)

#### 12. Research Objectives

(Align Research Objectives with Research Gaps, Research Questions, etc.)

(Research Objectives must be specific, measurable, achievable, realistic, time constrained) [S.M.A.R.T.]

#### 13. Resources/facilities accessible to execute the project

#### 14. Research Methodology

This section should clearly present each aspect of the process by which the study/research will be completed. Every attempt should be made to leave no question as to the procedures used to complete the study. Proper scientific methods should be used for this aspect of the study/research. Methods, Subjects, Instrumentation, Procedures, etc. (if relevant and applicable) Ensure that the research methodologies are appropriate for answering the research questions and that they are feasible within the available resources and time frame.

# 15. Feasibility Study of the Proposed Research Project Plan for planning and fruitful execution of Research Project.

#### 16. Describe Research Design (as applicable to the specific type of Research)

Correlational	Causal	Comparative	interviews
Quasi- Experimental	Experimental/ Laboratory	Simulation	surveys
Empirical	Meta-analytic	analytic	Participant observations
Applied	Basic/Fundamental	Qualitative/Creative	Oral history
Quantitative	Classification	Field	Archival research
Comparative	Source Criticism	Focus Groups	Case studies

(or any other type of research not covered above)

- **17. Sampling (if applicable)** describe the aspects of the cases on which data collection and analysis will focus (where relevant), Indicate how access to the study population will be achieved .
- **18.** Variables (As applicable to the Research Project with justification) describe aspects of the cases on which data collection and analysis will focus (where relevant).

Dependent	Quantitative	Latent
Independent	Qualitative	Continuous in
maependent	(categorical)	time
Control	Observable	Discrete time

#### 19. Methods of Data Collection (as applicable)

#### 20. Organization of Work Elements

Provide a timeline listing the order for all the major steps of the study and indicate the approximate amount of time needed for each step (Time schedule of activities giving major milestones, Time schedule of activities through Bar diagram)

#### 21. Data Analysis Procedures and Interpretation, if applicable:

Outline the data analysis methods (Qualitative as well as Quantitative) and how the results will be interpreted. Verify that the methods are appropriate for the data to be collected.

#### 22. Results/Findings:

Evaluate the presentation of results, including data tables, graphs, and figures. Verify if the results address the research questions, properly and if they are supported by the data collected.

#### 23. Conclusions, suggestions/recommendations:

List the conclusions drawn from the study and whether they are supported by the evidence presented in the project report.

#### 24. Discussion:

Check the interpretation of the results and the extent to which the findings align with the stated objectives. The discussion should include critical analysis and potential limitations.

#### 25. Future Research Directions/ Recommendations for further Research

Provide recommendations to further research on this topic or how parts of the study/research undertaken could be improved upon. If researcher found as a result of his/her study that another topic should be looked at in order to offer more insight into this topic, then he/she should suggest that at this time. It is important that this part of the conclusion chapter incorporates the implications of the findings drawn from the Research Project in terms of other research in the specific area of study, investigated by the researcher.

#### 26. Research Outcomes with beneficiaries

Outcome of research should result into Product Patent/ Process Patent/Design Patent and/or High quality Publications in Journals listed in UGC CARE List Group-I and UGC CARE List Group II. (Also list the possible beneficiaries of the research)

The outcome of the conducted research in PG programme is likely to be patented and/or publishable in the journals indexed in UGC CARE List Group-I and UGC CARE List Group-II.

#### 27. Sustainability in Research Project (if applicable, please specify)

- Human sustainability
- Social sustainability
- Economic sustainability
- Environmental sustainability (issues related to energy, Carbon footprint assessment, climate and biodiversity)
- Life Cycle Assessment (LCA). LCA enables the assessment of environmental impacts of a service or

product by taking into account all the stages of its life cycle according to different criteria, including but not limited to carbon dioxide CO2 measurement.

- Reproducibility of the protocols; results; research materials: product, information, data, software, Codes, etc.
- Use of Green artificial intelligence, which seeks to reconcile powerful computing with environment friendly research
- Adoption of Eco-friendly practices

# 28. Originality/Novelty of the project (Justify one or more of the following as applicable to the Research work)

a) Incremental improvement	b) Devise New investigative methods /analysis / synthesis
c) Substantial/radical improvement	d) Devise new system/model/product/process/ machine/ article of manufacture/composition of matter/new and useful improvement of any of these
e) Discover new information/model/system	f) Apply New Methods/Approach/Techniques/ Algorithms
g) Provide new Technical Solution to a Problem	h) Create New Interpretations
i) Modify existing theories/ systems/ models /Algorithms /Interpretations	j) Provide additional support for existing theories/models/interpretations
k) Analyze phenomena/Results of Research in new ways	l) Disprove the existing theories/ models/ interpretation
m) Generation of New Data	n) Devise new original and ornamental design for an article of manufacture
o) Invention or discovery and reproduction of any distinct and new variety of plant (Botany)	p) Any other aspect not covered above

### 29. Plagiarism/Similarity Check

This requires that the researcher's work:

- Provides a full and complete representation of any scholarly findings
- Credits the contributions of other researchers, colleagues, co-workers, etc.
- Respects diversity of opinion

#### Misconduct in research and writing is defined as

- The fabrication, falsification, plagiarism, or other practices that seriously deviate from those commonly accepted within the scientific, artistic, and academic professional communities.
- Plagiarism involves the intentional appropriation of another's work, including ideas or phrasing of words, without crediting the source.
- Please include the similarity analysis report or plagiarism check report of the Entire Research Project by Urkund (or Turnitin or iThenticate or any other software available at the Knowledge Resource Centre of Sant Gadge Baba Amravati University, Amravati).
- This will ensure the originality of the Research work. UGC's new anti-plagiarism policy allows up to 10% content similarity. With similarity above it, students will be asked to revise and resubmit the synopsis.

[Reference: University Grants Commission (Promotion of Academic Integrity and Prevention of Plagiarism in Higher Educational Institutions) Regulations, 2018.]

- 30. Prior Approval from the IAEC registered with the CPCSEA (The Committee for Control and Supervision of Experiments on Animals) to be submitted to the Ph.D. Cell along with the Research Synopsis (If Applicable to the Research Work)
  - As per the "Breeding of and Experiments on Animals (Control and Supervision) Rules, 1998", an Institutional Animal Ethics Committee (IAEC) is needed for control and supervision of experiments on animals performed in the Institute/Research Laboratory. The IAEC must be registered with the CPCSEA.
  - Prior approval of the IAEC is mandatory for all types of research proposals involving small animal experimentation before the start of the study. This Committee also monitors research throughout the study and after completion of the study through periodic reports besides regular visits to the research faculty animal house and laboratories where the experiments are conducted. This also ensures compliance with all regulatory requirements, rules, guidelines and laws related to animal experiments.
  - A copy of the prior approval of the IAEC, which is already registered with the CPCSEA must be enclosed along with the Research Synopsis

### 31. References/Bibliography

- Credibility of Sources of literature: Journals used in the literature review must be indexed in the UGC- CARE Group-II or UGCCARE List Group-I.
- References/ bibliographies must be listed in uniform standard style (APA/ MLA/Chicago/IEEE/MHRA or Harvard)
- Ensure that all references are properly referred to in the running text and Seminal Research Articles are included in the references.

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